



# Development of a conjugate vaccines for enteric fever

8th International Conference on Typhoid Fever and Other Invasive Salmonellosis

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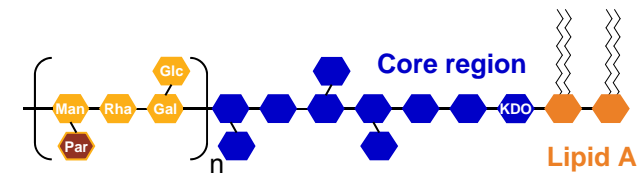
# NVGH enteric fever vaccine strategy

## *Bivalent vaccine against S. Typhi and S. Paratyphi A*

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- Glycoconjugate combination vaccine
  - Building on NVGH development of Vi-CRM<sub>197</sub>
  - Employing Novartis know-how and expertise

- S. Paratyphi A component
  - Serovar specific O-antigen, O:2
  - Covalently linked to CRM<sub>197</sub>



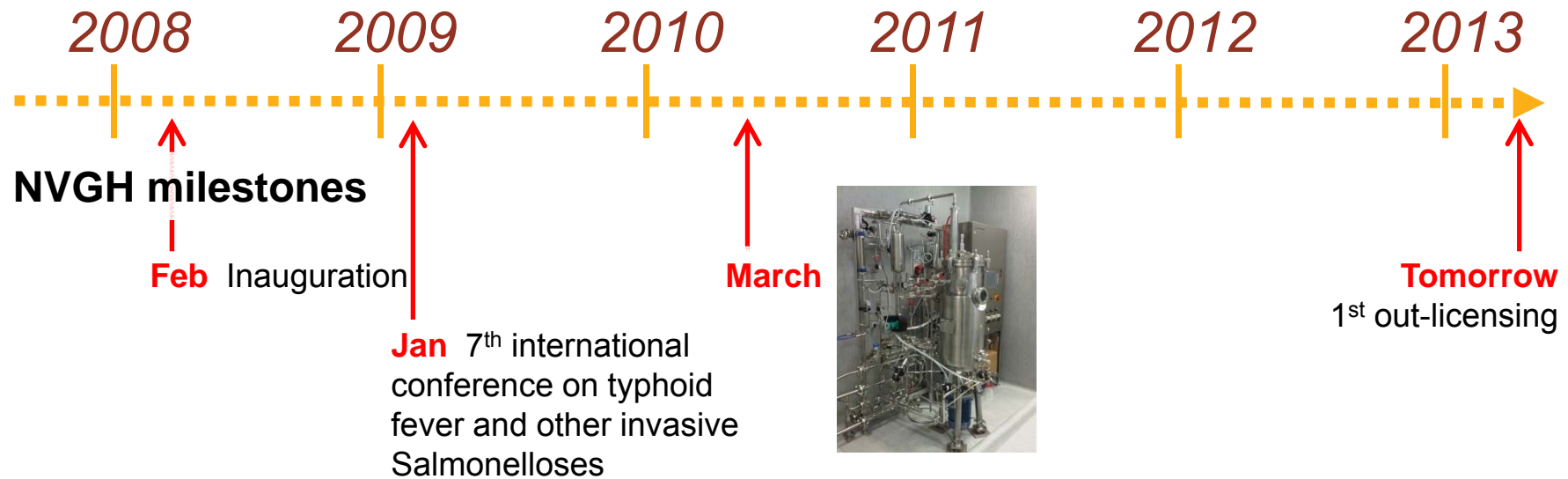
### O:2 of S. Paratyphi A

External portion of Lipopolysaccharide (LPS)

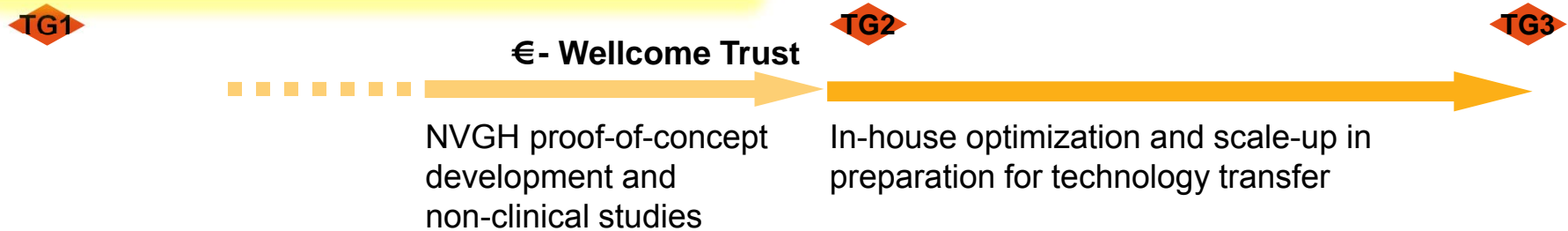
- Bivalent vaccine
  - VI-CRM<sub>197</sub> + O:2-CRM<sub>197</sub>

# Bivalent vaccine development since Kilifi Kenya

*Building on Vi-CRM<sub>197</sub> and other NVGH projects*



## O:2-CRM<sub>197</sub> and bivalent highlights



# NVGH enteric fever vaccine, bivalent conjugate

## Unique attributes

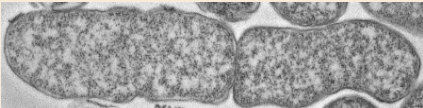
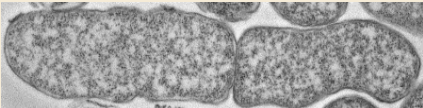
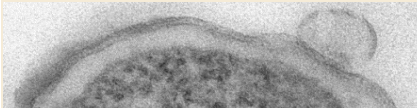
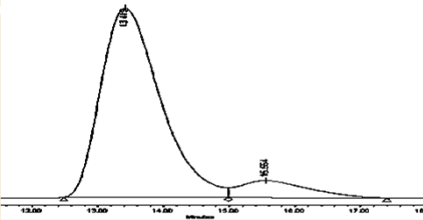
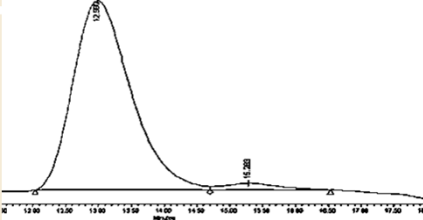
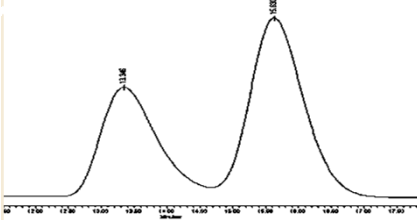
Micoli et al, PLoS One 2012 & Micoli et al, Anal Biochem 2013

- Uses readily available, scalable GMP materials and equipment
- O:2-CRM<sub>197</sub> designed to be compatible with Vi-CRM<sub>197</sub>

Process area	O:2-CRM <sub>197</sub>
<b>O:2 source</b> MOGM S. Paratyphi A	Drug sensitive for safety Genetically modified to produce more membrane Grows well in defined, simple media
<b>O:2 purification</b> No column chromatography	Efficient <i>in situ</i> extraction method No hazardous or expensive reagents or intermediates
<b>Carrier protein</b> CRM <sub>197</sub>	Mutant diphtheria toxin (used by Novartis Vaccines and Diagnostics)
<b>Conjugation</b> O:2 per CRM <sub>197</sub> < 2	Novel method developed by NVGH
<b>Cost of Goods</b>	Similar to Vi-CRM <sub>197</sub>
<b>Bivalent formulation</b>	No overt interactions observed

# O:2 polysaccharide production source

*Wild-type/attenuated vs GMMA producing line*

Characterization	Wild-type	Attenuated	GMMA producing
Genetic modifications	none	$\Delta$ guaBA	$\Delta$ ToIR
Morphology			
O:2 hydrolysis	biomass	biomass	biomass + GMMA
O:2 heterogeneity HMW : MMW	70 : 30	80 : 20	10 : 90
			
Average repeating units	~ 45	< 55	~ 25
% O-acetylation	~ 70	> 70	~ 50

- MMW O:2 easier to handle gives, more consistent conjugates

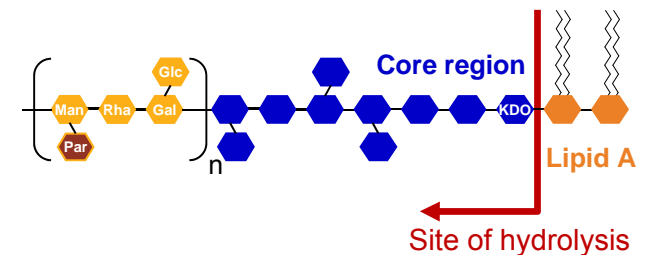
# O:2 polysaccharide production process

Optimized for 30 L scale

Micoli et al, Anal Biochem 2013

- Shake flask
- High cell density fermentation (30 L)
- O:2 antigen hydrolysis in situ (100°C, > 4 h)
- Neutralization in situ
- Harvest (TFF Microfiltration)
- Ultrafiltration (TFF 30 kDa cut-off)
- Precipitation 1 (pH 3)
- Centrifugation
- Negative chromatography (Sartobind S)
- Precipitation 2 (EtOH + CaCl<sub>2</sub>)
- Centrifugation
- Ultrafiltration (TFF 10 kDa cut-off)
- Filtration (0.2 µm)
- **Purified O:2 antigen Intermediate (Bulk)**

→ Exploiting sterilize in place vessel



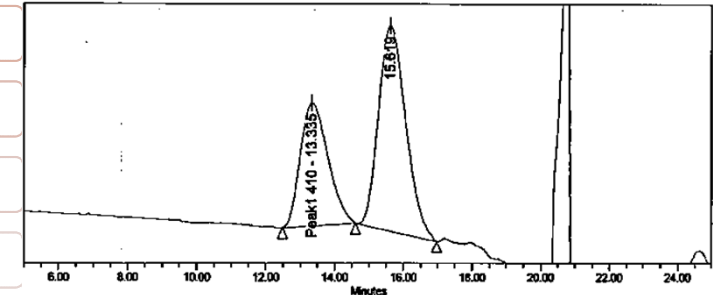
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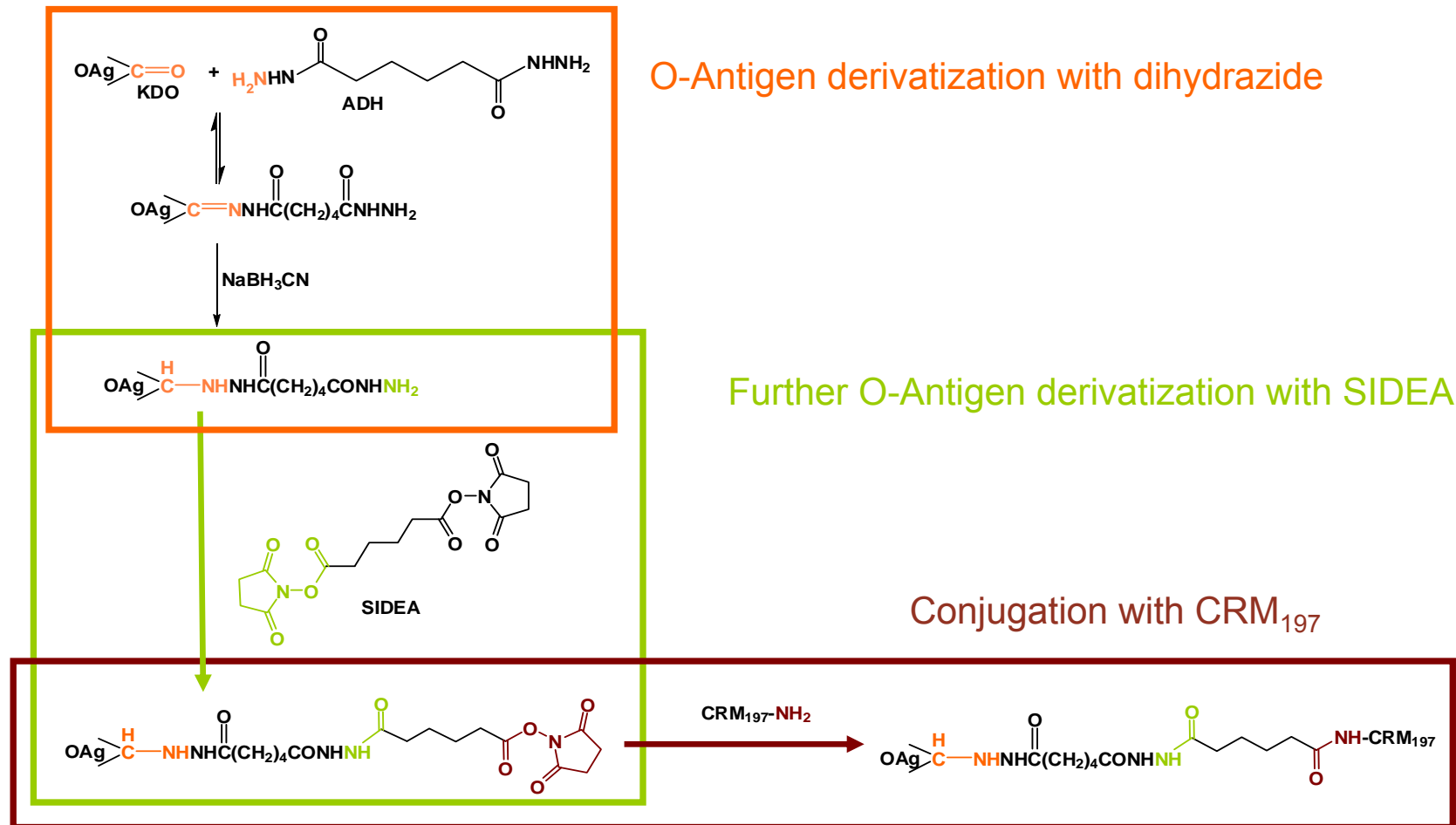
- O-acetylation > 40 %
- Protein < 0.5 %
- Nucleic acid < 0.5 %
- Endotoxin < 0.01 UI/µg

→ Good purity & well characterized

# O:2-CRM<sub>197</sub> conjugation process

Also used in the iNTS conjugate vaccines

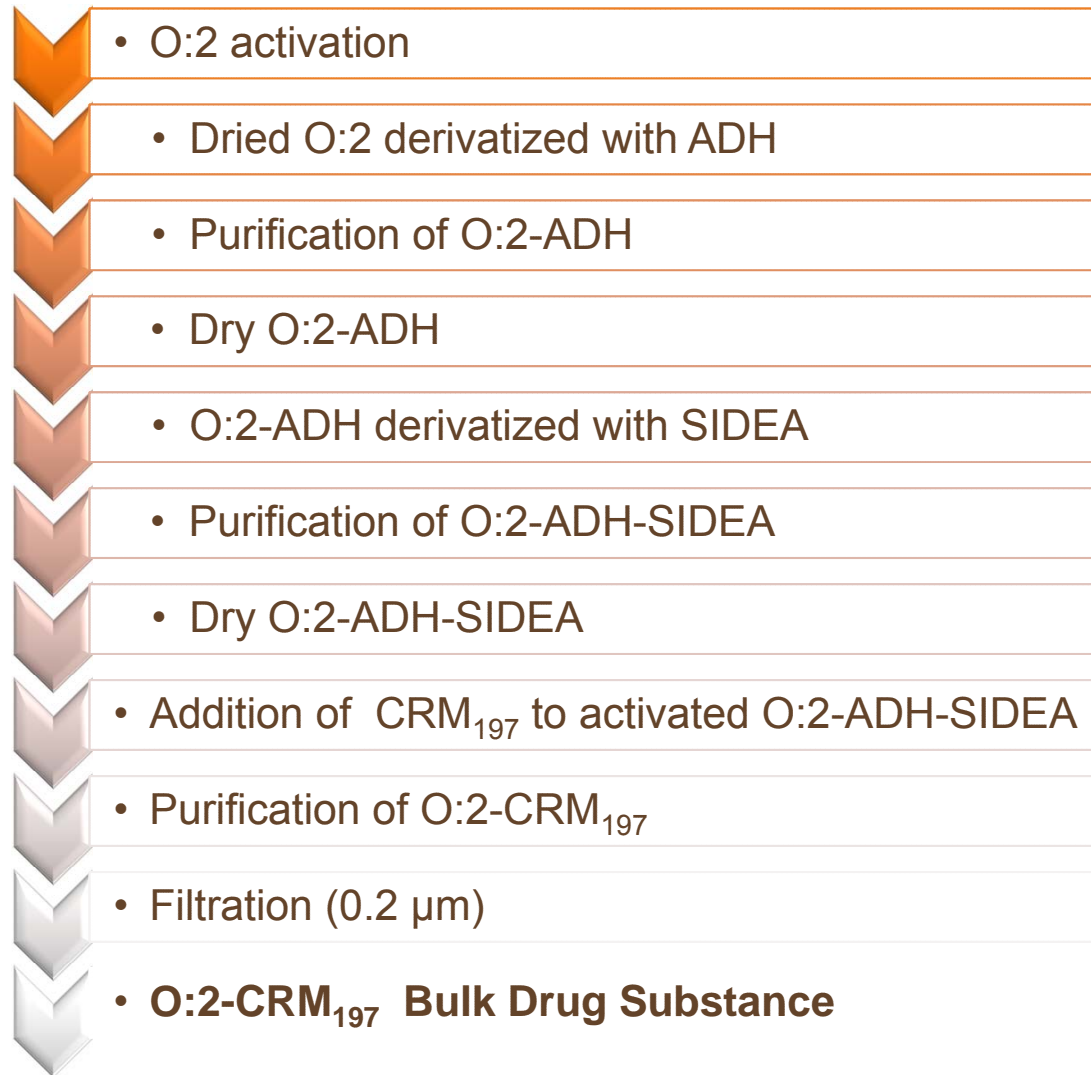
Micoli et al, PLoS One 2012



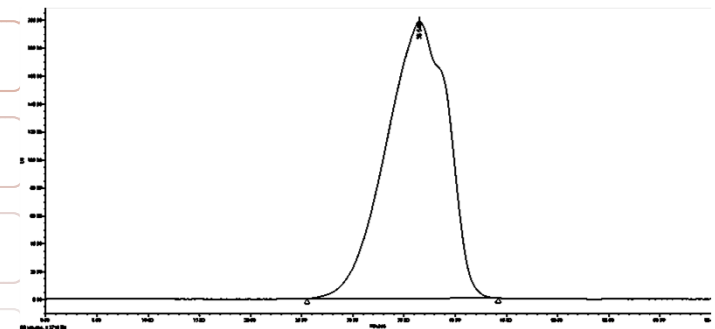


# O:2-CRM<sub>197</sub> production process

Optimized for 200 mg scale



→ **Selective chemistry through KDO, terminal sugar, of core**

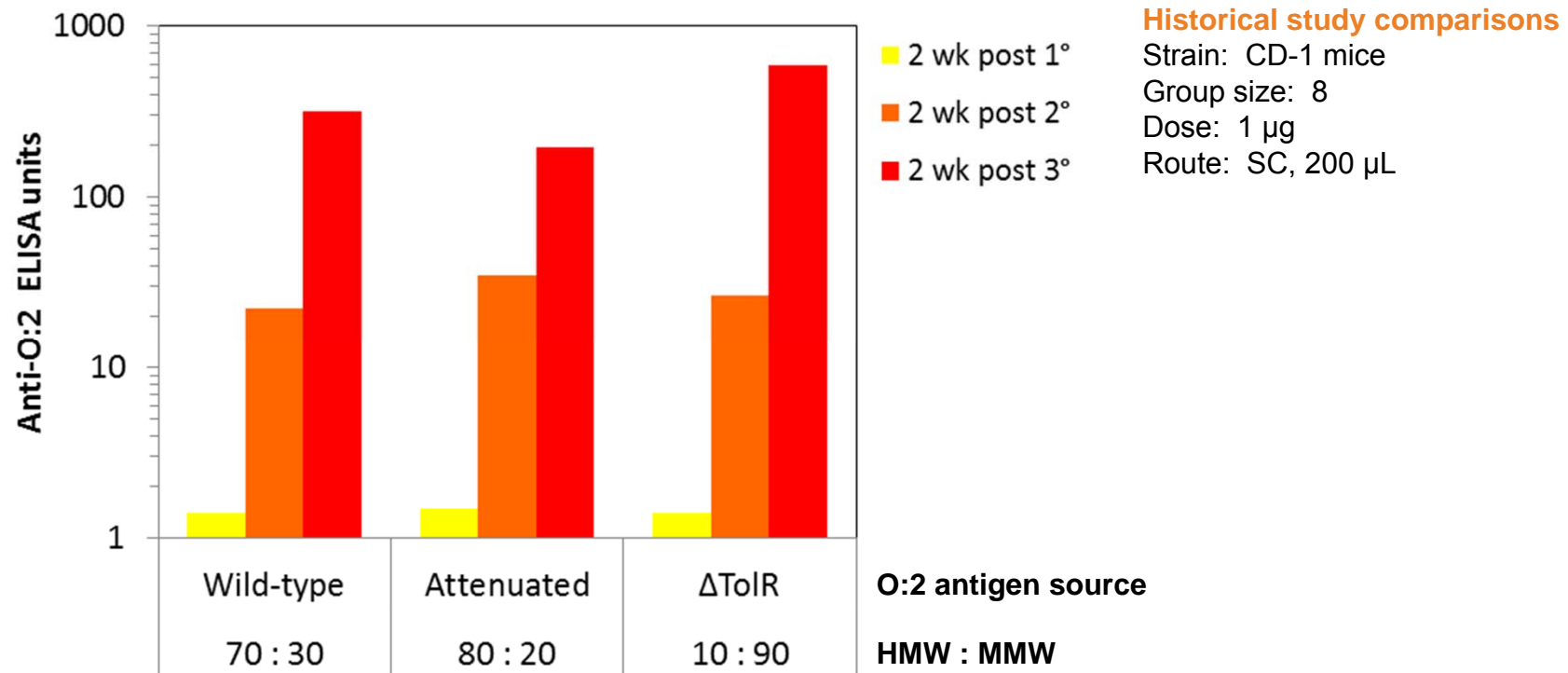


- PS : Protein 1.3
- Yield CRM<sub>197</sub> 80 %
- O-acetylation > 40 %

→ **Good purity & well characterized**

# O:2-CRM<sub>197</sub> immunogenicity

Similar responses from wild-type, attenuated and  $\Delta$ ToIR strains

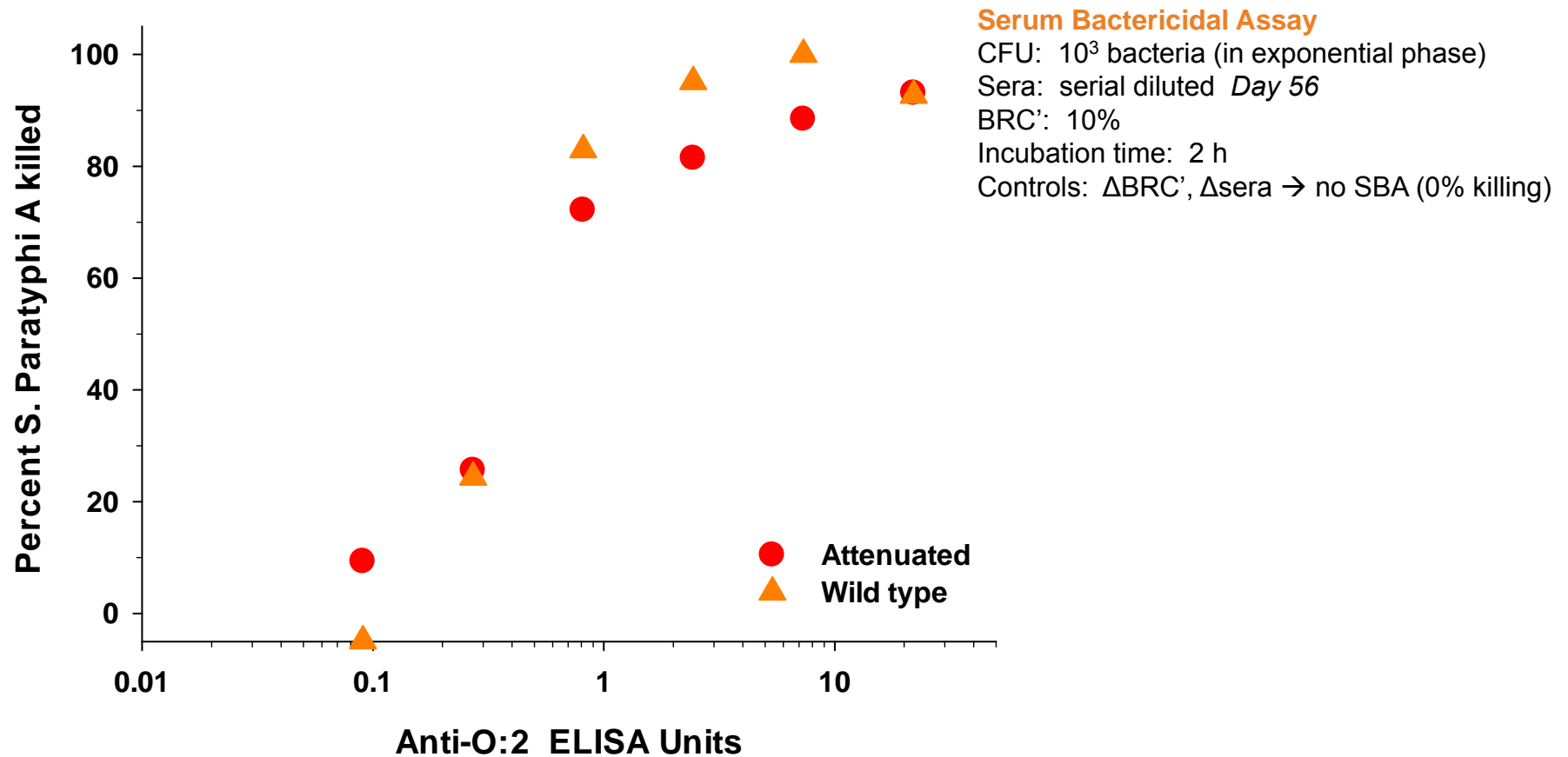


Anti-O:2 serum IgG responses are not highly dependent on

- Source of O:2 antigen
- Ratio of the MW populations in the O:2
- Level of O-acetylation

# O:2-CRM<sub>197</sub> induces functional antibodies

*Increasing antibody = increased killing*



- O:2-CRM<sub>197</sub> produces antibodies that can kill *S. Paratyphi A*
- Impact of combining Vi-CRM<sub>197</sub> with O:2-CRM<sub>197</sub> . . .

# Bivalent vaccine antibodies are bactericidal

## Against both Vi+ bacteria (*Citrobacter*) and *S. Paratyphi A*

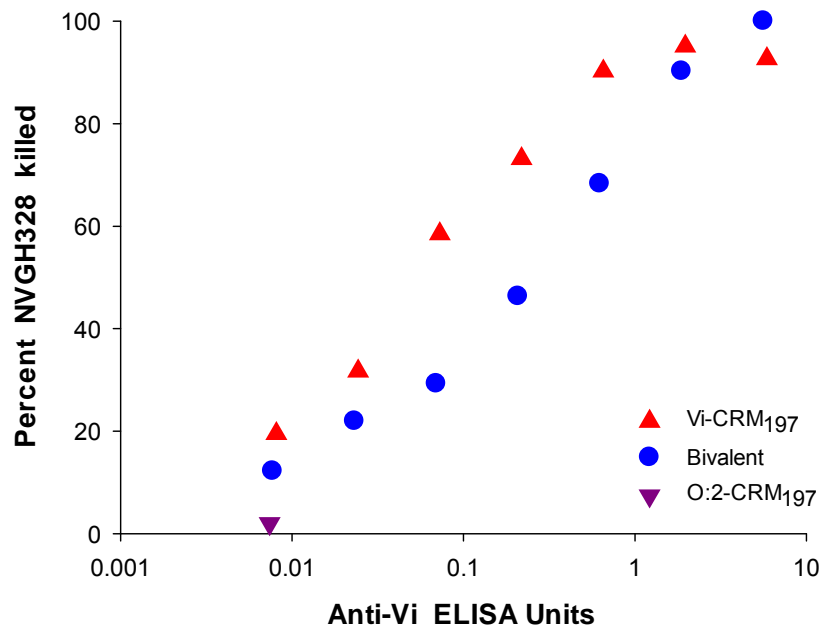
### Study Design

Strain: CD-1 mice                      Route: SC, 200  $\mu$ L  
 Group size: 8                          Immunization: days 1, 14 & 42  
 Dose: 1  $\mu$ g antigen                      Bled: day 56  
 Vaccines: Vi-CRM<sub>197</sub>, O:2-CRM<sub>197</sub> or bivalent  
**ELISA results: no immunologic interference**

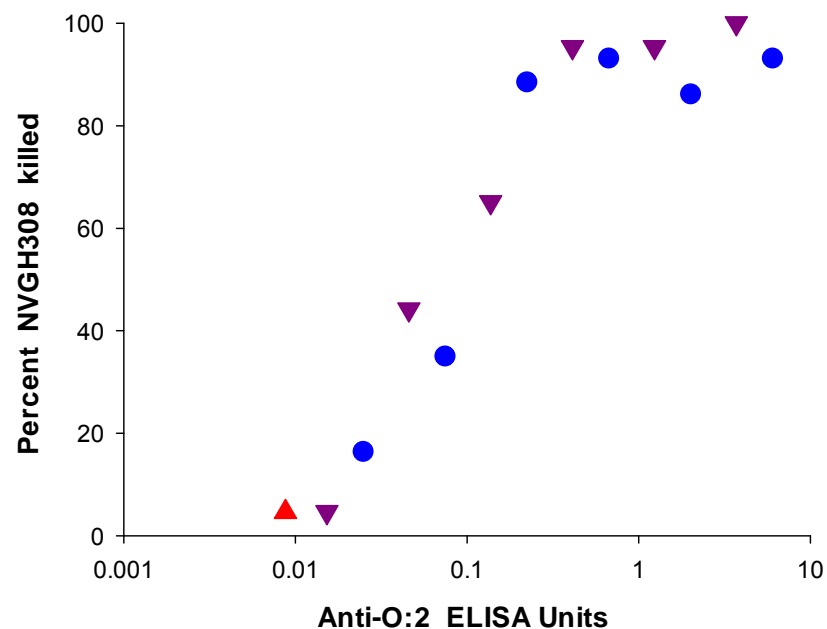
### Serum Bactericidal Assay

CFU: 10<sup>3</sup> bacteria (in exponential phase)  
 Sera: serial diluted Day 56  
 BRC': 10%  
 Incubation time: 2 h  
 Controls:  $\Delta$ BRC',  $\Delta$ sera  $\rightarrow$  no SBA (100% survival)

***Citrobacter* killing correlates with anti-Vi levels**



***S. Paratyphi A* killing correlates with anti-O:2 levels**



- Vi-CRM<sub>197</sub> + O:2-CRM<sub>197</sub> likely to provide coverage against enteric fever

# NVGH enteric fever vaccine, what is next

## *Reducing the risk and meeting the mission*

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- Proof of principle for a bivalent, Typhoid / Paratyphoid A, vaccine
  - Technical and animal immunogenicity
  
- Activities for 2013 and beyond
  - Prepared to transfer the robust processes to a commercialization partner
    - Vi-CRM<sub>197</sub> for typhoid fever
    - Bivalent (Vi-CRM<sub>197</sub> + O:2-CRM<sub>197</sub>) for enteric fever
  - Support partner for further technical and clinical development
  
- Aiming for developing country access to enteric fever vaccine
  - Initial registration and roll out in India
  - WHO prequalification and wider distribution throughout S. Asia

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## *Working together with collaborators and contributors*

### **NVGH Enteric Fever Development Project Teams**

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Think what is possible