

**Attenuated non-typhoidal  
*Salmonella* strains as live oral  
vaccines and as reagent strains for  
conjugate vaccine production**

**Sharon Tennant**

Center for Vaccine Development (CVD),  
University of Maryland School of Medicine  
Baltimore, MD, USA

# A vaccine against invasive NTS

## *Target age by disease burden*

- Sub-Saharan Africa
  - Children < 36 months of age
  - Deliver through the Expanded Program on Immunization (EPI) along with pentavalent vaccine, OPV and rotavirus vaccine at ~ 6, 10 & 14 weeks of age
- North America, Europe
  - Elderly
  - Deliver along with future *Clostridium difficile* and norovirus vaccines, as well as influenza and future elderly pneumococcal vaccine

# Roles for CVD attenuated NTS strains

- As live oral vaccines (*guaBA clpPX*)
- To make **conjugate vaccine** (COPS-FliC) production safer & more economical:

$\Delta$ *guaBA*- Primary attenuating mutation. Bacteria require exogenous guanine for growth in vitro. This deletion allows large-scale fermentation with **enhanced occupational health safety**

$\Delta$ *clpPX*- Secondary attenuating mutation results in enhanced flagella expression for **economical purification** of flagella

$\Delta$ *fliD*- Flagellin exported as monomers which enables **economical purification** of flagellin



# Engineered NTS strains constructed

## *Prototype strains (for proof of principle in mice)*

### **S. Typhimurium I77 (ST19)**

- CVD 1921 --  $\Delta guaBA \Delta clpP$  (hyperexpresses flagella)
- CVD 1925 --  $\Delta guaBA \Delta clpP \Delta fliD \Delta fljB$  (hyperexpresses Phase 1, but not Phase 2, flagellin monomers)

### **S. Enteritidis R11**

- CVD 1941 --  $\Delta guaBA \Delta clpP$
- CVD 1943 --  $\Delta guaBA \Delta clpP \Delta fliD$

## *Definitive oral vaccine strains (for clinical trials)*

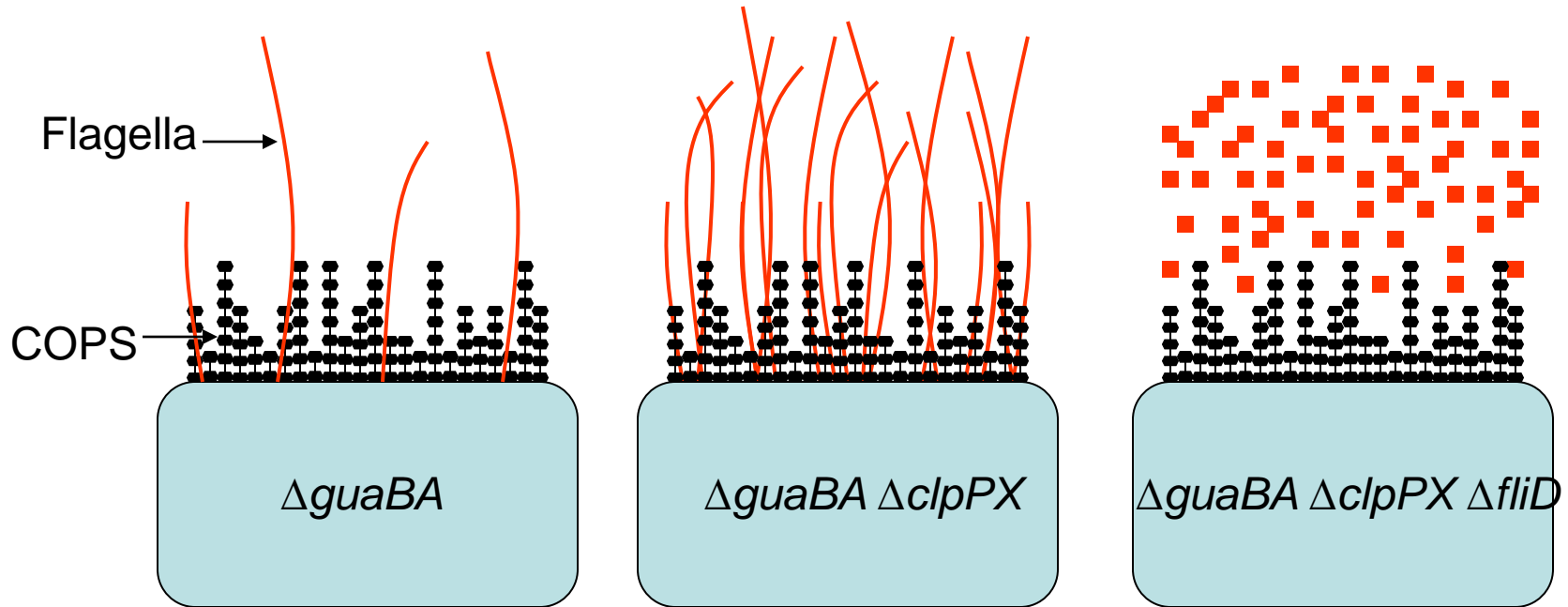
### **S. Typhimurium D65 (ST313)**

- CVD 1931 --  $\Delta guaBA \Delta clpX$

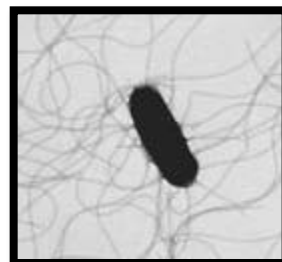
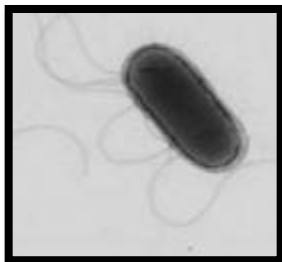
### **S. Enteritidis R11**

- CVD 1944 --  $\Delta guaBA \Delta clpX$

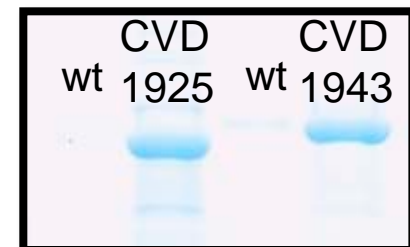
# Phenotypes of attenuated strains



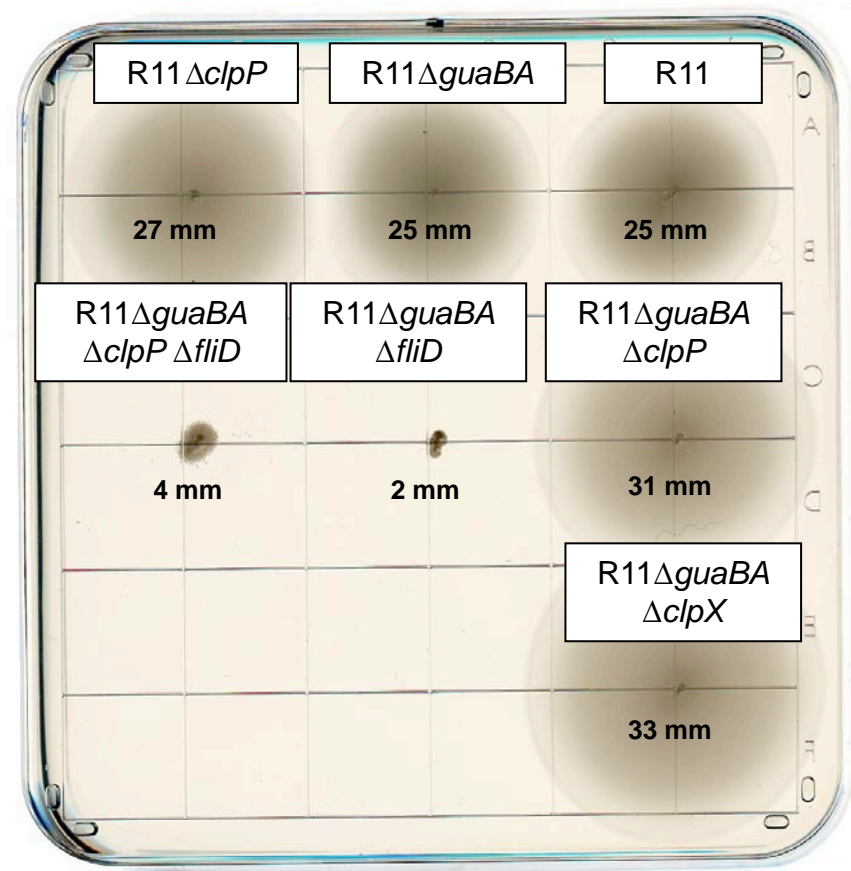
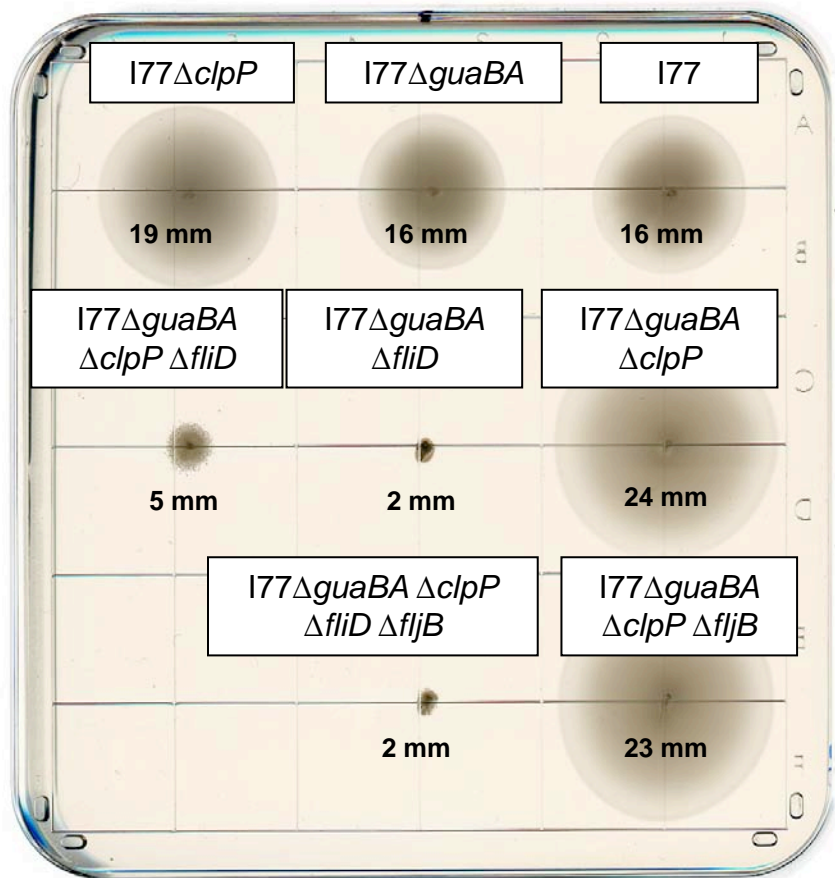
LIVE  
VACCINE



REAGENT  
STRAIN



# NTS *clpP*X mutants are more motile than wild-type and *fliD* mutants are non-motile

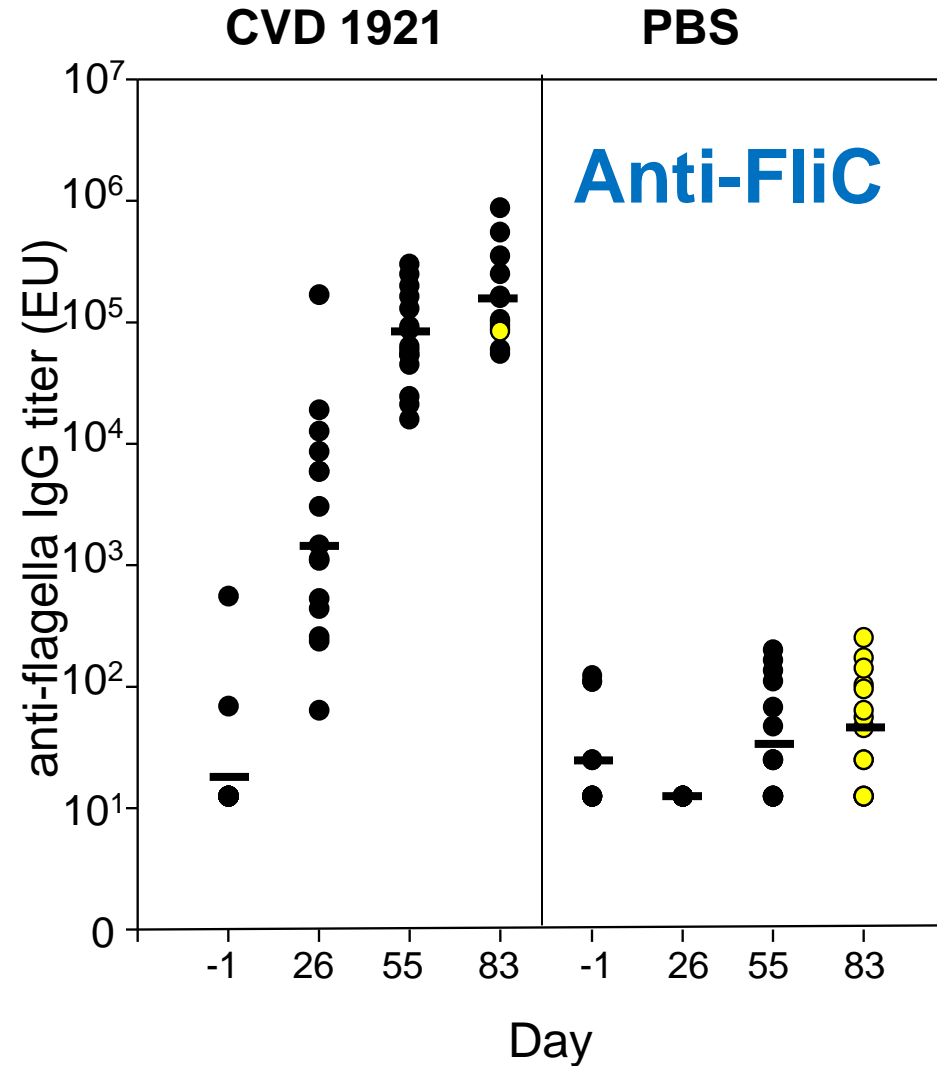
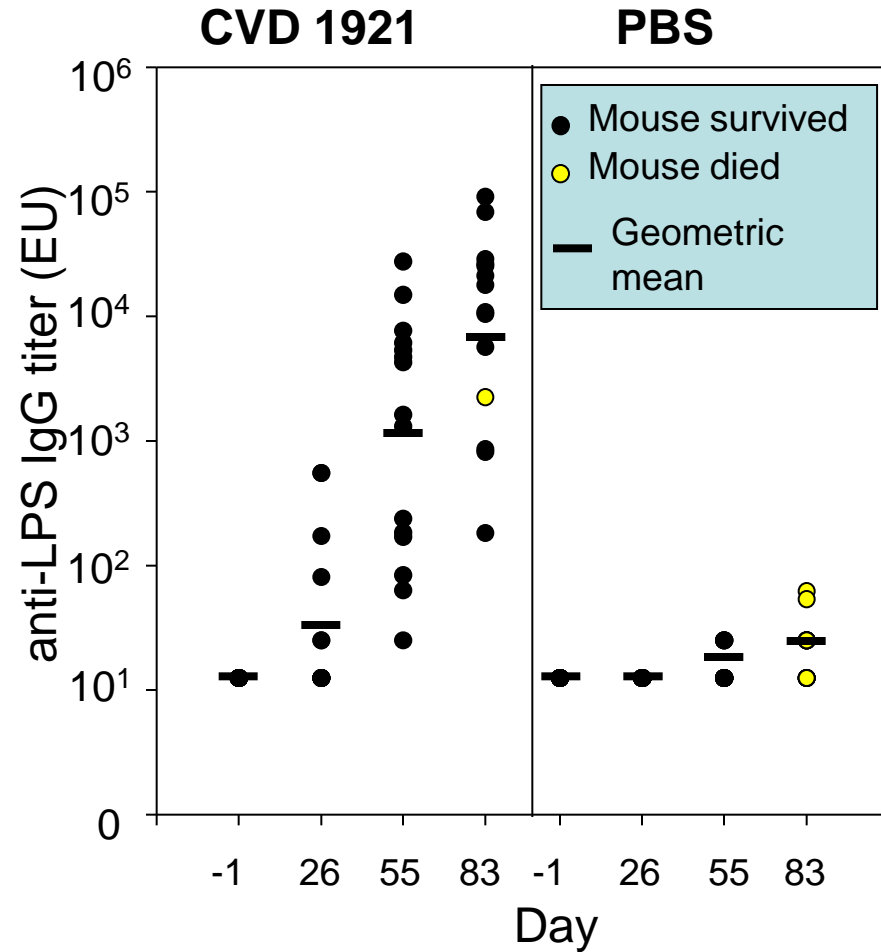


# NTS strains harboring $\Delta guaBA$ , $\Delta clpP$ and deletions are highly attenuated in mice

| Strain                      | LD50 <sub>p.o.</sub> |
|-----------------------------|----------------------|
| Wild-type                   | $2 \times 10^4$ CFU  |
| $\Delta guaBA$              | $>10^9$ CFU          |
| $\Delta guaBA, \Delta clpP$ | $>10^{10}$ CFU       |

# Immunized mice produce high serum IgG anti-LPS and anti-FlaC titers

## Anti-LPS



Mice were immunized on days 0, 28 and 56 and challenged on day 84 with 100 LD<sub>50</sub>'s.



# Live attenuated *S. Typhimurium* vaccines mediate homologous but not heterologous protection

| Vaccine                                      | Challenge                                  | Challenge dose | Vaccine efficacy | P value |
|--|--|----------------|------------------|---------|
| CVD 1921<br>( <i>ΔguaBA ΔclpP</i> )<br>ST19  | <i>S. Typhimurium</i> I77<br>(B; i; 1,2)   | 100 X LD50s    | 86%              | P<0.001 |
| CVD 1931<br>( <i>ΔguaBA ΔclpX</i> )<br>ST313 | <i>S. Typhimurium</i> D65<br>(B; i; 1,2)   | 10,000 X LD50s | 100%             | P<0.001 |
| CVD 1931<br>( <i>ΔguaBA ΔclpX</i> )<br>ST313 | <i>S. Stanleyville</i> J65*<br>(B; z4,z23) | 3 X LD50s      | 91%              | P<0.001 |
| CVD 1931<br>( <i>ΔguaBA ΔclpX</i> )<br>ST313 | <i>S. Enteritidis</i> R11<br>(D; gm)       | 50 X LD50s     | <b>51%</b>       | P=0.07  |

Mice were immunized orally on days 0, 28 and 56 and challenged orally on day 84

\* i.p. challenge

# Live attenuated *S. Enteritidis* vaccines mediate homologous and heterologous protection

| Vaccine                             | Challenge                                | Challenge dose | Vaccine efficacy | P value |
|-------------------------------------|--|----------------|------------------|---------|
| CVD 1941<br>( <i>ΔguaBA ΔclpP</i> ) | <i>S. Enteritidis</i> R11<br>(D; gm)     | 100 X LD50s    | 76%              | P<0.001 |
| CVD 1944<br>( <i>ΔguaBA ΔclpX</i> ) | <i>S. Enteritidis</i> R11<br>(D; gm)     | 10,000 X LD50s | 83%              | P<0.001 |
| CVD 1944<br>( <i>ΔguaBA ΔclpX</i> ) | <i>S. Dublin</i> R17<br>(D; gp)          | ~800 X LD50s   | 85%              | P<0.001 |
| CVD 1944<br>( <i>ΔguaBA ΔclpX</i> ) | <i>S. Typhimurium</i> D65<br>(B; i; 1,2) | ~200X LD50s    | 81%              | P=0.002 |

Mice were immunized orally on days 0, 28 and 56 and challenged orally on day 84

# Live oral vaccine summary 1

- Target populations for NTS vaccines include high risk groups for mortality in the USA (elderly) & Africa (infants & perhaps HIV-positive adults)
- Candidate live oral vaccines and reagent strains have been constructed
- Prototype  $\Delta guaBA \Delta clpPX$  attenuated NTS strains are:
  - Safe, immunogenic and protective in mice
  - Safe in SIV-infected Rhesus macaques

## Live oral vaccine summary 2

- Live NTS vaccines elicit significant seroconversion (4-fold or > rise) of anti-LPS & anti-flagellin antibody titers
- Antibodies show functional antibody activity
- Definitive live oral NTS vaccines have been shown to be protective against a highly lethal challenge in mice
- A live attenuated *S. Enteritidis* vaccine was able to mediate cross-protection against *S. Typhimurium* but not vice versa

# Acknowledgements

**CVD**



**Center for Vaccine Development (CVD)**

**University of Maryland Baltimore**

Jin Y. Wang

Patrick Schmidlein

Deanna Toema

Girish Ramachandran

Mary Boyd

Raphael Simon

James E. Galen

Marcela Pasetti

Sofie Livio

Orit Gat

Myron M. Levine

This work was funded by the Middle Atlantic RCE Program, NIAID/NIH 2 U54 AI057168-06.