

Multidrug-Resistant *Salmonella enterica* in the Democratic Republic of the Congo (DRC)

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Burden of disease – Sub Saharan Africa

***Salmonella* Typhi:**

- 77.4 / 100.000 (children + adults) (Buckle 2012)
- Case fatality rate of 1% (Crump2004)

Non-typhi *Salmonella* (NTS):

- 175-388 / 100.000 (children) (Gordon2012)
- 1800-9000 / 100.000 (non-ART, HIV⁺ adults) (Gordon2012)
- Case fatality rate of 22%-25% (children + adults) (Gordon 2008)

=> No incidence data from Central Africa available



Reported resistance rates – Central Africa

Salmonella Typhi

- Low numbers of MDR, increasing to >50% since mid 90's
- Very low rates of Fluorquinolone resistance
- No resistance to 3rd gen. Cephalosporin

(Vlieghe, 2009)

Non-typhi Salmonella

- High numbers of resistance to Ampicillin and Chloramphenicol
- Medium resistance to Cotrimoxazole and Fluorquinolones
- No resistance to 3rd gen. Cephalosporins
- Resistance to Cotrimoxazol + Fluorquinolones observed from 1999 onwards

(Vlieghe, 2009)



Trigger for Survey

- In 2004 / 2005 an outbreak of *Salmonella* Typhi was observed in Kinshasa
- Case fatality rates of >50% were observed (Muyembe-Tamfum, 2008)
- All isolates evaluated (n=11) were MDR but susceptible to:
 - Gentamicin
 - Ciprofloxacin
 - Cefotaxim

=> A project to assess current susceptibility status of *Salmonella* spp. in the DRC was implemented



Methods – Surveillance

From **2007 to 2011** a prospective health care facility based passive survey at centers in **7/11 provinces**:

- Inclusion criteria: suspicion of invasive bacteremia
- Standard demographic data was recorded
- Blood for culture was collected
 - Standard laboratory procedures + antisera testing performed at Institut National de Recherche Biomédicale, (Kinshasa, DRC)
 - Re-serotyping + AB susceptibility testing performed at the Institute of Tropical Medicine (Antwerpen, Belgium)
 - PFGE + molec. markers for fluoroquinolone resistance performed at the National Institute of Public Health (Brussels, Belgium)



Methods – Antimicrobial Susceptibility

- Antitibiotic susceptibility testing for **ampicilin, cefotaxime, trimethoprim-sulphamethaxole (TMP-SMX)** was performed using the Vitek II (bioMérieux)
- **MIC** for nalidixic acid, ciprofloxacin, chloramphenicol and azithromycin was determined using E-test macromethod (bioMérieux)
- **ESBL** testing was done with double disc diffusion method (CLSIM100S22)



Definitions

- Minimal inhibitory concentrations for **nalidixic acid** ($\text{MIC} \geq 32$ mg/l) and **chloramphenicol** ($\text{MIC} \geq 16$ mg/l) were determined according to CLSIM100S21
- **Azithromycine** resistance: $\text{MIC} > 16$ mg/l (EUCAST v2.0)
- Decreased ciprofloxacin susceptibility (**DCS**): $\text{MIC} > 0.064$ mg/l (EUCAST v2.0)
- Multi Drug resistance (**MDR**): Resistance against first line antibiotics ampicillin, chloramphenicol, cotrimoxazol (TMP-SMX)



Methods – Molecular Analysis

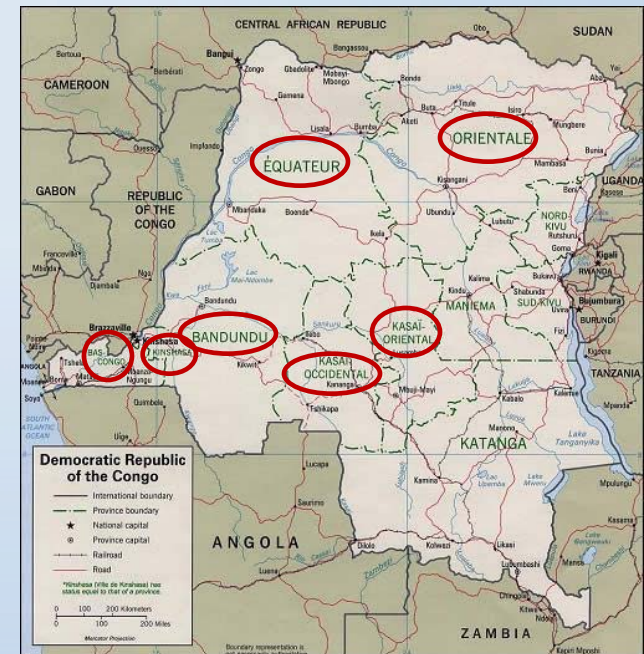
- **Pulsed field gel electrophoresis (PFGE)** was performed on a subset using *XbaI* as restriction enzyme according to PulseNet protocol
- **Screening** for chromosomal quinolone resistance determining regions (QRDR): *gyrA*, *gyrB*, *parC* genes (CEQ2000 DNA sequencer, Beckman Coulter)
- **Screening** for plasmid mediated quinolone resistance genes (*qnrA*, *qnrB*, *qnrS*) (Cavaco, 2009)



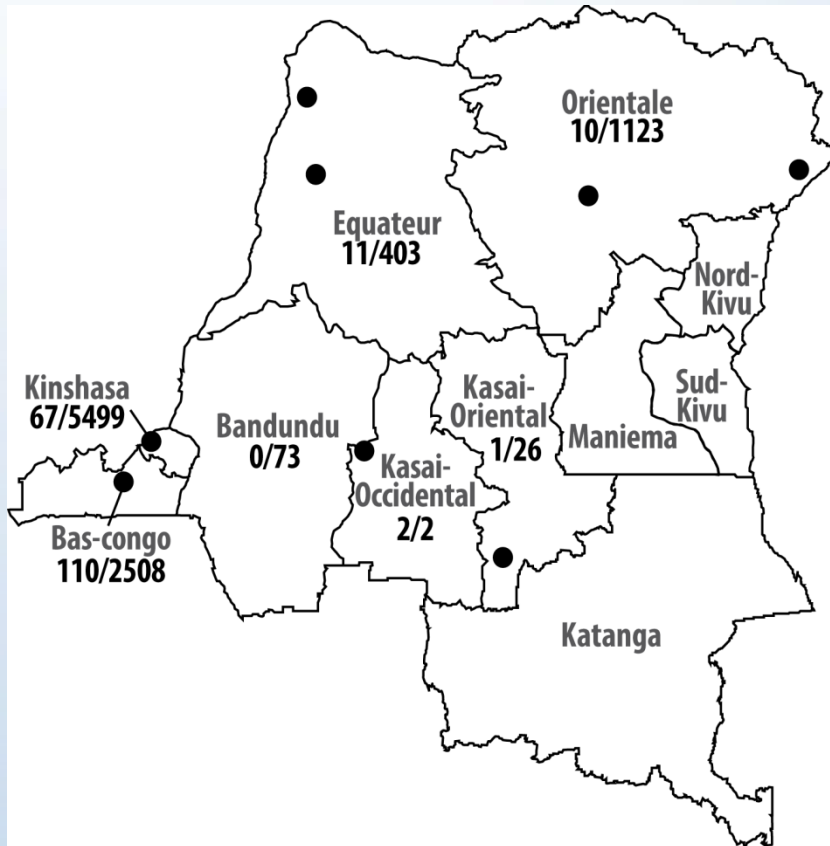
Results

A total of **9.634 blood** samples were collected in 7/11 provinces and cultured

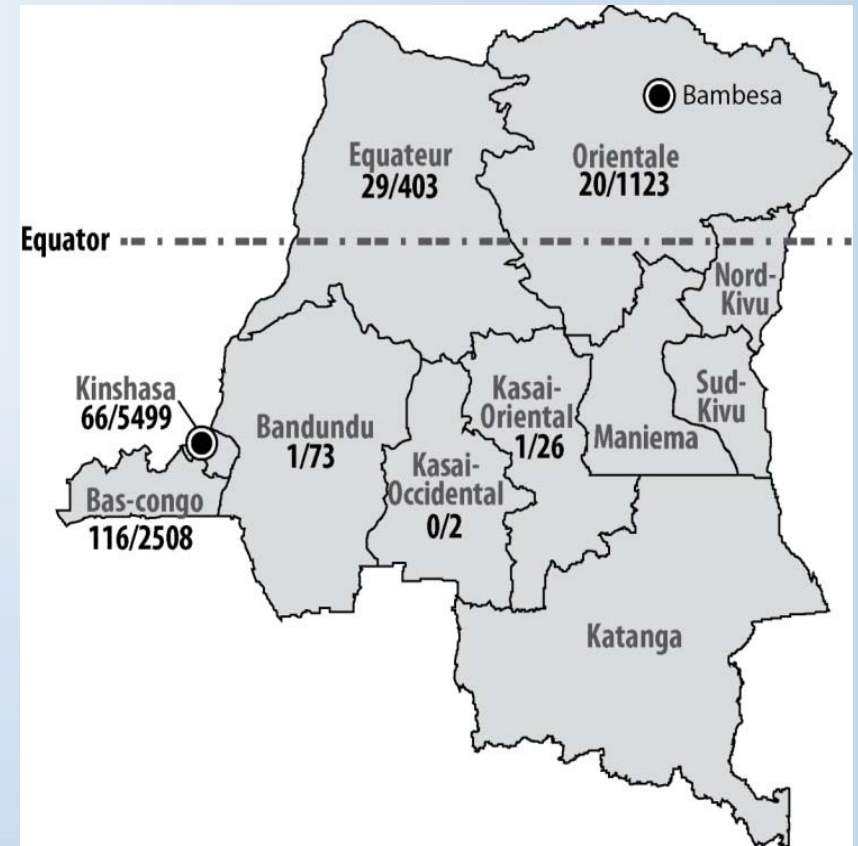
- Positivity rate: 989 (10.3%, excluding contaminants)
- *Salmonella* Typhi: 201 (20.3%)
- NTS: 233 (23.6%)
 - 184 *Salmonella* Typhimurium (79%)
 - 42 *Salmonella* Enteritidis (18%)
 - 7 other *Salmonella* spp. (3%)



Salmonella Typhi



Non – typhi *Salmonella*



Results – Antimicrobial Resistance (%)

	<i>Salmonella</i> Typhi (n=201)	non-typhi <i>Salmonella</i> (n=233)	<i>Salmonella</i> Typhimurium (n=184)	<i>Salmonella</i> Enteritidis (n=42)
Ampicillin (%)	64.7	88.0	94.0	64.3
Chloramphenicol (%)	41.3	83.7	90.2	61.9
TMP-SMX (%)	57.7	88.0	94.0	64.3
MDR (%)	30.3	80.7	86.9	59.5
DCS (%)	15.4	4.3	4.9	0.0
Nalidixic Acid (%)	15.4	4.3	4.9	0.0
MDR+DCS (%)	7.5	3.9	4.3	0.0
Azithromycin (%)	1.0	3.0	3.3	0.0
Cefotaxime (%)	0.0	2.1	2.2	0.0
MDR+DCS+ESBL (%)	0.5	0.9	0.5	0.0



Mechanisms of DCS

Salmonella Typhi (n=31, all DCS):

- A total of 31 DCS associated point mutations:
 - *gyrA*: 83Ser > Tyr or Phe (n=22)
 - *gyrA*: 87Asp > Gly or Tyr or Asn (n=9)
- No *qnrA* and *qnrB* genes were detected
- No mutations in *gyrB* and *parC* genes



Mechanisms of DCS

Non-typhi *Salmonella* (n=10, all DCS):

- *gyrA*: 87Asp> Tyr (n=8)
- *gyrA*: 87Asp>Asn (n=2)
 - Also ESBL producers (all type SHV)
- No *qnrA*, *qnrB* and *qnrS* genes were detected
- No mutations in *gyrB* and *parC* genes



Pulsed Field Gel Electrophoresis

- ***Salmonella Typhi*** (n=185): 30 Profiles detected:
 - 132 (71%) shared one profile
 - Main profile over time and space
 - 41 (31%) were MDR
 - 23 (17%) were DCS
 - 11 (8%) were MDR + DCS
- ***Salmonella Typhimurium*** (n=34): 19 Profiles detected:
 - 7 (21%) shared one profile (T4)
- ***Salmonella Enteritidis*** (n=16): 10 Profiles detected:
 - 4 (25%) shared one profile (E5)



Discussion – *Salmonella* Typhi

MDR and DCS were observed for *Salmonella* Typhi

- MDR was less frequent as had been reported in previous studies from the region
- DCS was more frequent as had been reported earlier from the region and was associated with point mutations in *gyrA*
- We possibly observed emerging azithromycine resistance



Discussion non-typhi *Salmonella*

- Very high rates of MDR were observed for **NTS**
- Resistance to 3rd generation cephalosporins + ESBL in NTS is reported for the first time from the DRC
- MDR rates in *Salmonella* Enteritidis were significantly lower than in *Salmonella* Typhimurium ($p < 0.01$)



Conclusion

- The observed rates of MDR and DCS underline the importance of permanent antibiotic stewardship programs in the DRC.
- The appearance of strains resistant to 3rd generation cephalosporins and azithromycin may be an indicator to spreading resistance against these drugs
- Comprehensive surveillance systems and public health interventions targeting *Salmonella* spp. are urgently needed to reduce the high burden of disease.
- Incidence studies on burden of disease are planned.





**Thank you
very much!**

