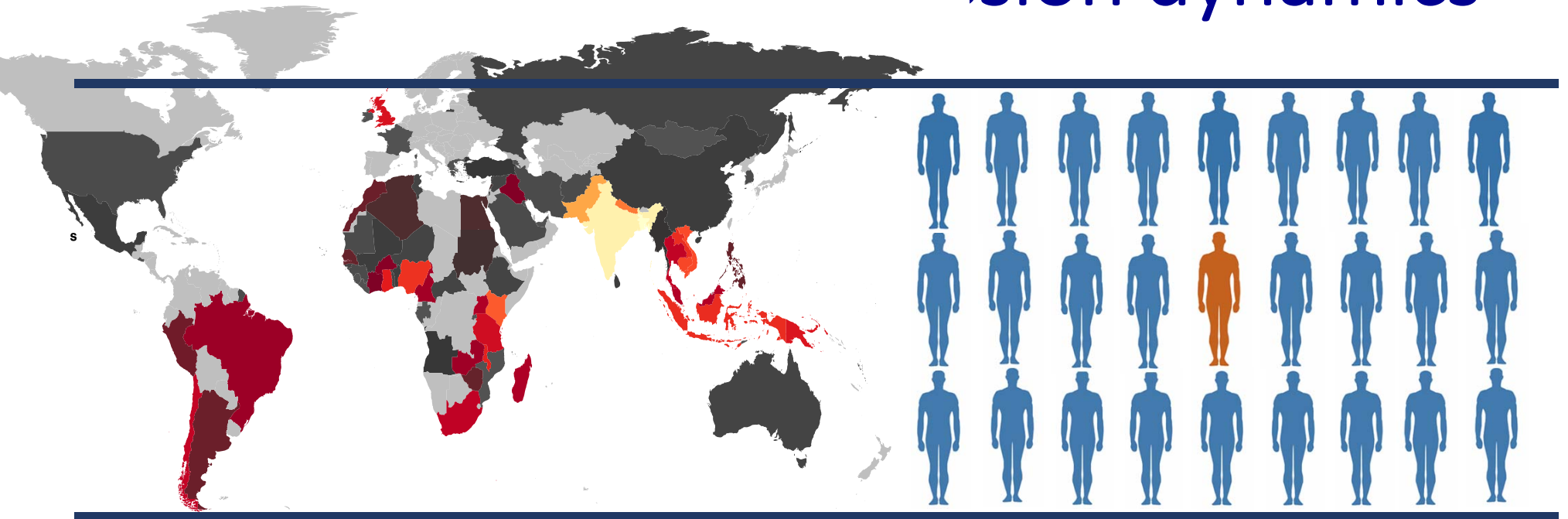


The role of genomics in typhoid control: sentinel traveler surveillance, in-host evolution and transmission dynamics



Dr. Zoe Anne Dyson
University of Cambridge & University of Melbourne

 [@msmicrobiocode](https://twitter.com/msmicrobiocode)
zad24@medschl.cam.ac.uk

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Acknowledgements

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Marie Chattaway

Satheesh Nair

Philip Ashton

Hassan Hartman

Joanne Freedman

Martin Day

Christian Medical College, India

Balaji Veeraraghavan

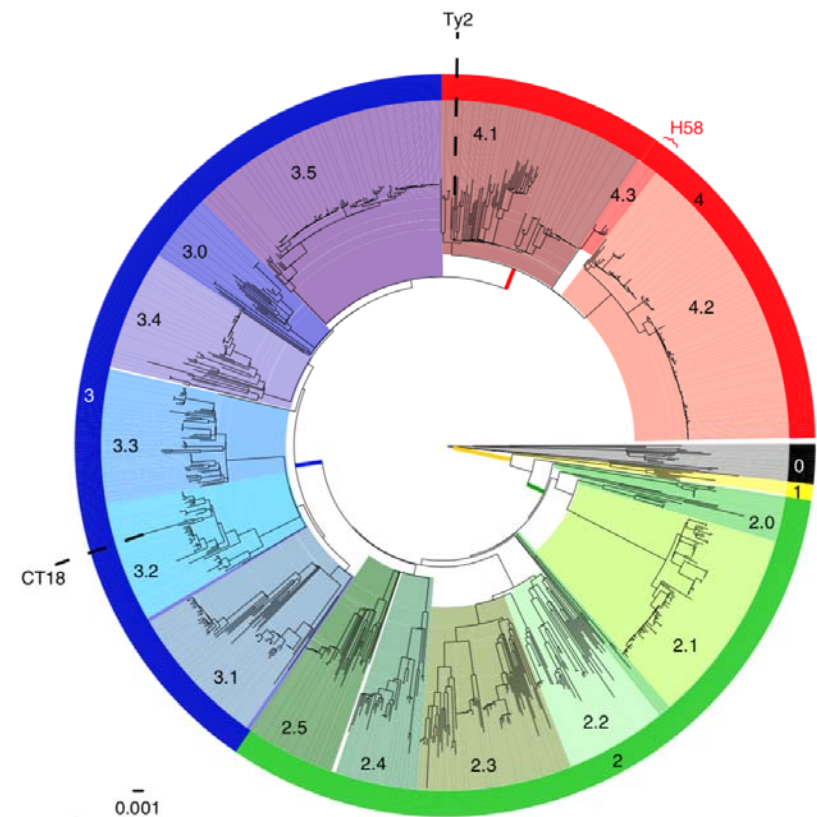
**St. Johns Medical College Hospital &
St. Johns Research Institute, India**

Savitha Nagaraj



Global genomic framework of *S. Typhi*

- **Global framework of ~2000 *S. Typhi***
 - Highly structured population
 - Strong geographical clustering
 - GenoTyphi tool: calls genotypes from genomes



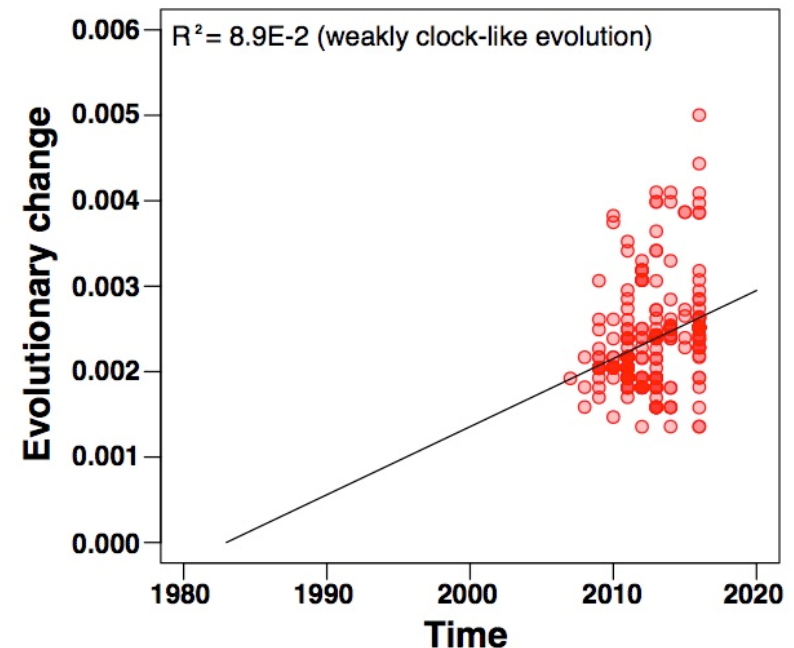
Global population structure of *Salmonella Typhi* (49 subclades)

Source: Wong *et al.* 2016, Nat. Commun.

 <https://github.com/katholt/genotyphi>

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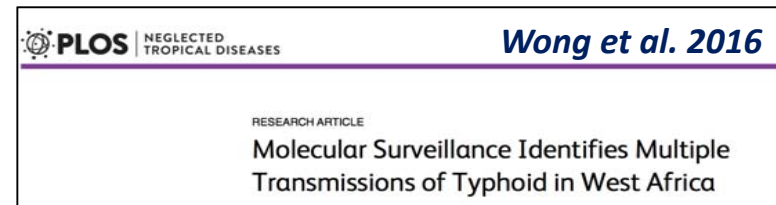
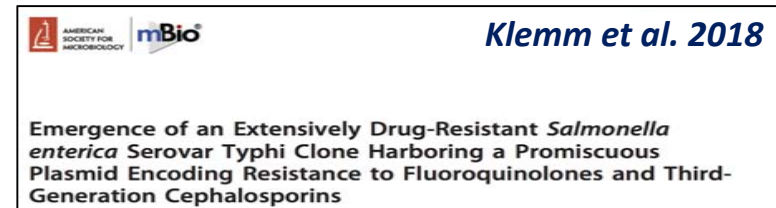
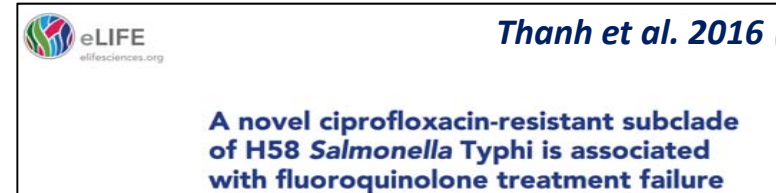
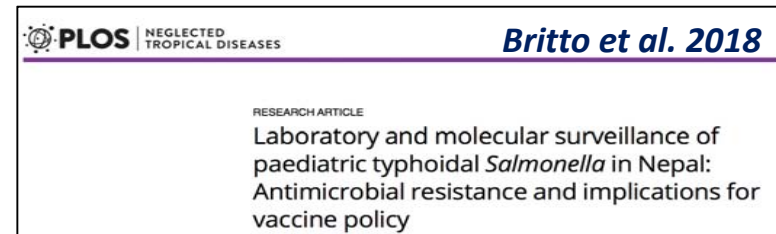
Weak temporal signal in Nepal H58 (4.3.1) Typhi

Source: Britto *et al.* 2018, PLoS NTDs

Wong *et al.* 2015, Nat. Genet.

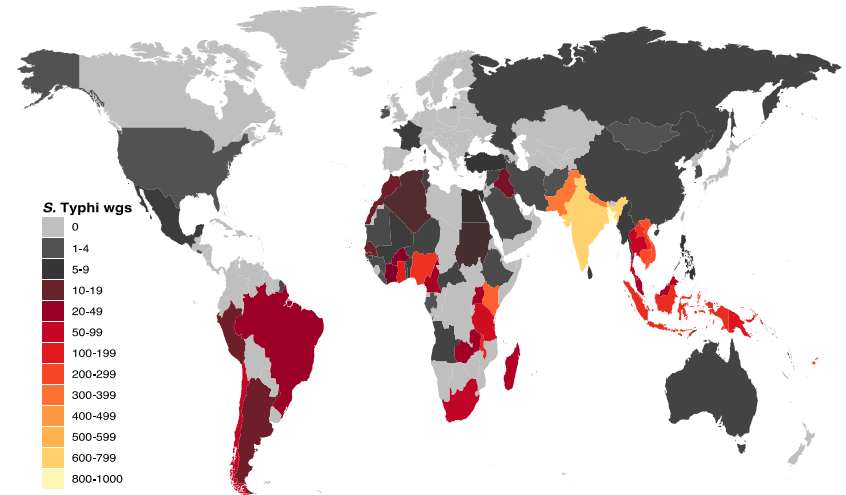
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 - A slow substitution rate of ~1 SNP/genome/year
- **Global genomic framework provides context for local epidemiological studies**
 - AMR, circulating genotypes, transmission dynamics



Open questions in typhoid genomics

1. Where are the gaps in typhoid WGS (whole genome sequencing) surveillance and can travel-associated typhoid cases help fill these?



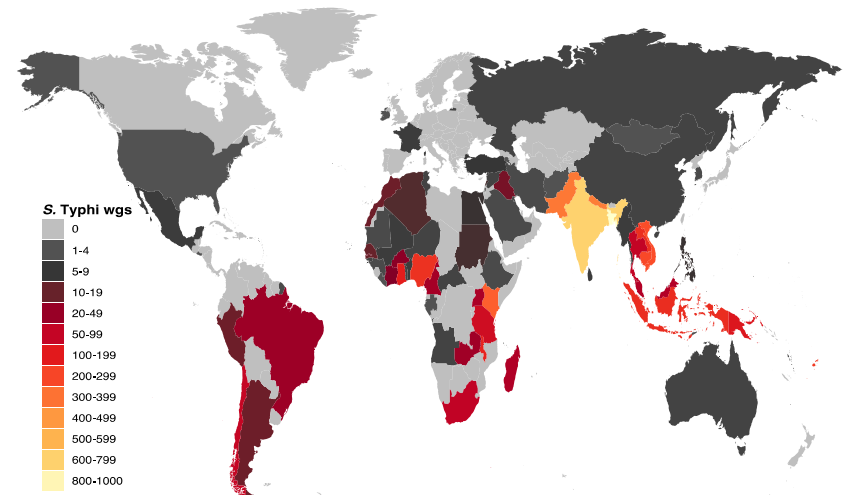
Location of travel for 533 *S. Typhi* WGS from Public Health England (PHE) from 2014-2017

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Please see: Poster 108 “*Salmonella* Genomics: A Revolution in Public Health Microbiology”, Nair *et al.* PHE

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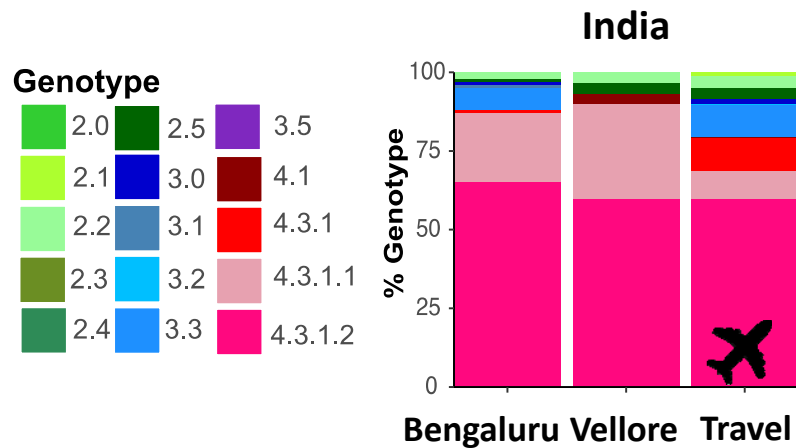
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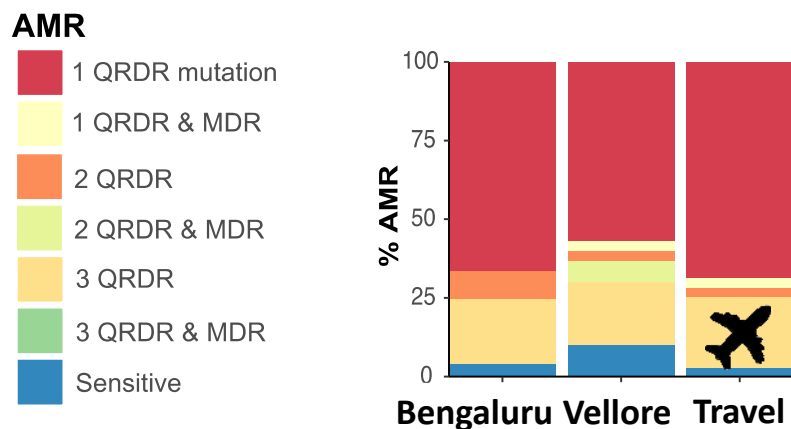
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Travel-associated *S. Typhi* are useful sentinels for surveillance in endemic countries

Genotypes

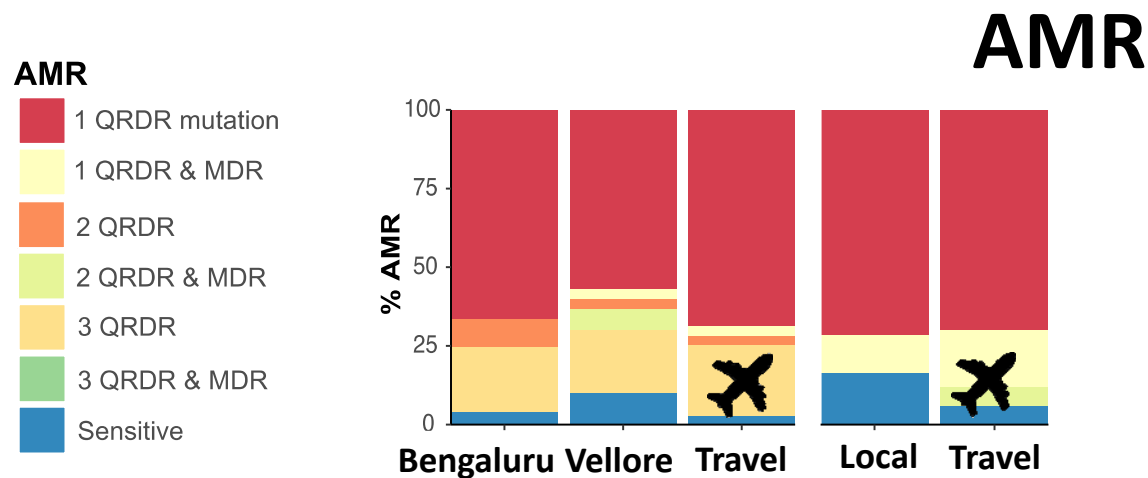
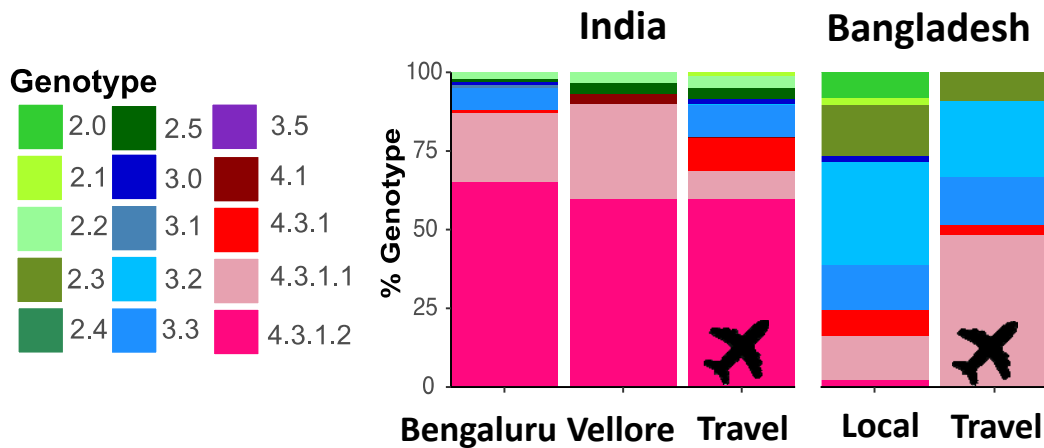


AMR



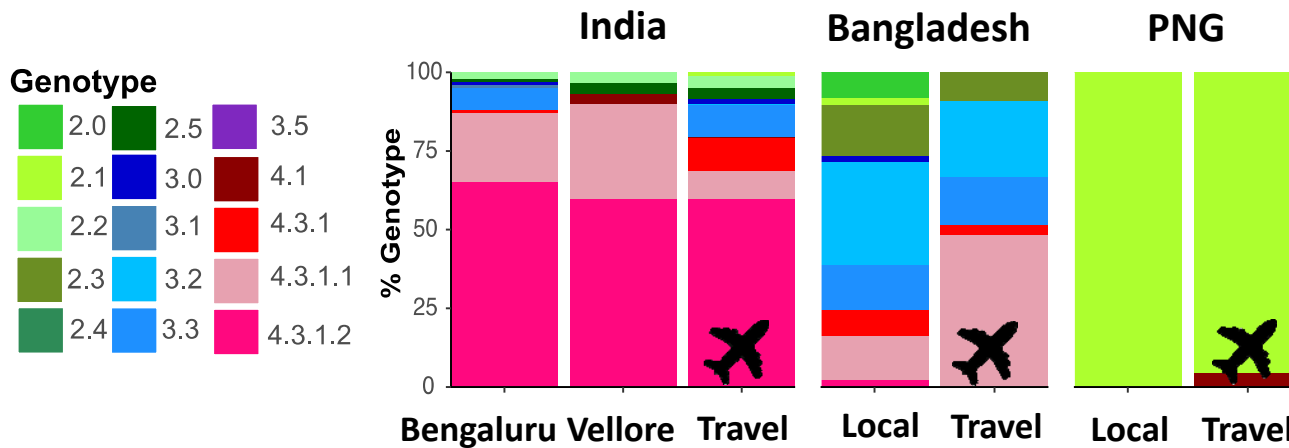
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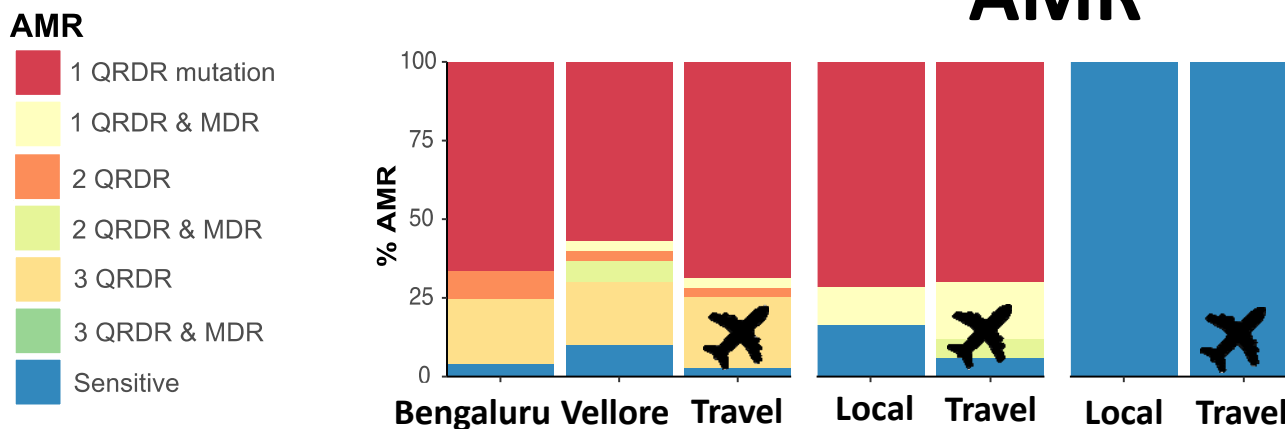


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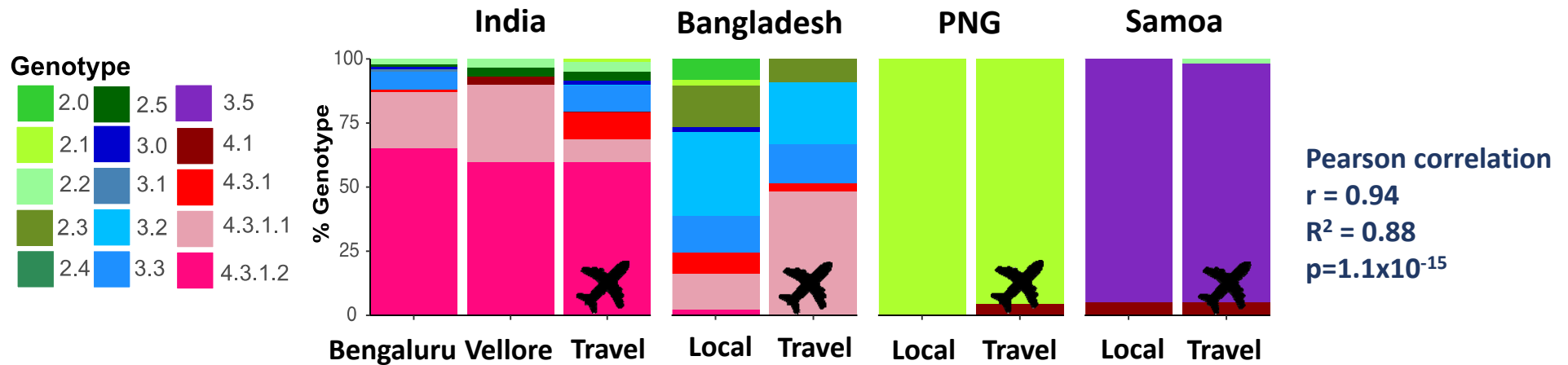


AMR

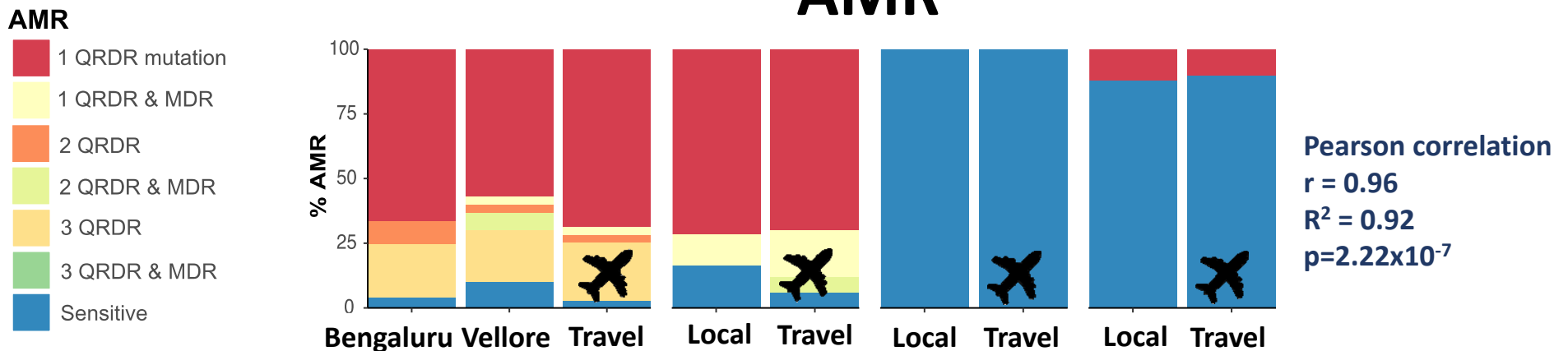


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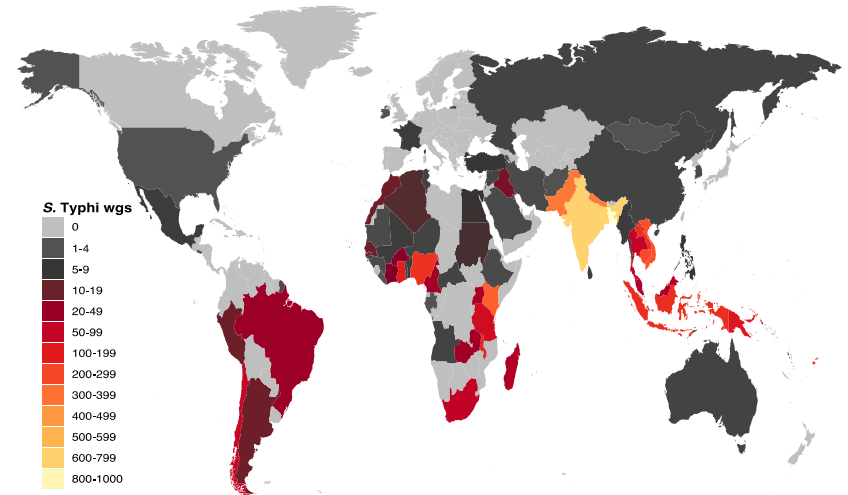


AMR



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In vivo evolutionary rate & transmission

RESEARCH ARTICLE

When are pathogen genome sequences informative of transmission events?

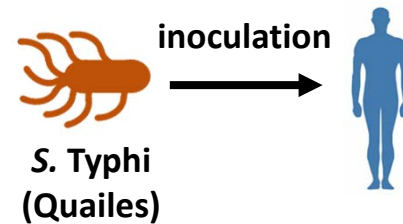
Finlay Campbell^{1*}, Camilla Strang², Neil Ferguson¹, Anne Cori^{1*}, Thibaut Jombart^{1*}

Table 1. Epidemiological and genomic parameters for ten major outbreak causing pathogens.

Pathogen	Generation time (SD) (in days)	Mutation rate (per site per day)	Genome length (base pairs)	Basic reproduction number R_0
<i>EBOV</i>	14.4 (8.9)	0.31×10^{-5}	18958	1.8
<i>MERS-CoV</i>	10.7 (6.0)	0.25×10^{-5}	30115	1.2
<i>SARS-CoV</i>	8.7 (3.6)	1.14×10^{-5}	29714	2.7
<i>Influenza A (H1N1)</i>	3.0 (1.5)	1.19×10^{-5}	13155	1.5
<i>MRSA</i>	15.6 (10.0)	5.21×10^{-9}	2842618	1.3
<i>K. pneumoniae</i>	62.7 (24.0)	6.30×10^{-9}	5305677	2.0
<i>S. pneumoniae</i>	6.6 (1.8)	5.44×10^{-9}	2126652	1.4
<i>M. tuberculosis</i>	324.4 (384.5)	0.24×10^{-9}	4411621	1.8
<i>S. sonnei</i>	8.5 (3.0)	1.64×10^{-9}	4825265	1.1
<i>C. difficile</i>	28.4 (14.9)	0.88×10^{-9}	4290252	1.5

Human challenge model of infection

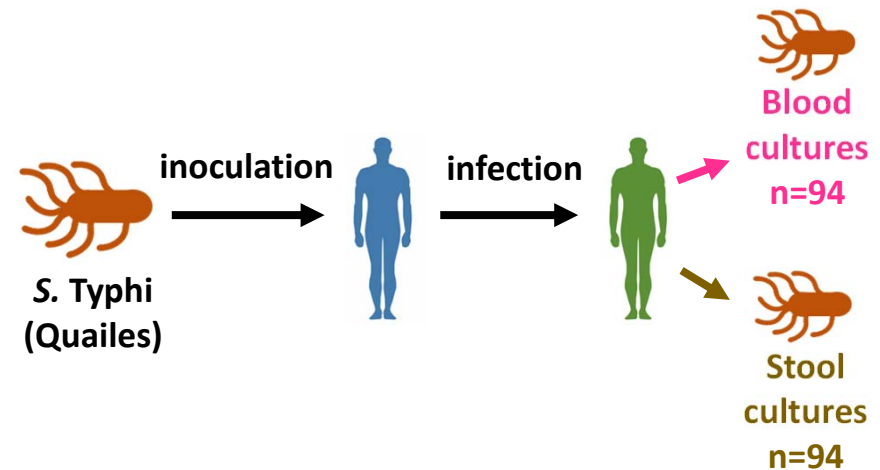
- 189 post challenge stool and blood cultures from 4 studies
- Compared WGS data from post-challenge strains to the inoculating (Quailes) strain



n=38, TI. Observational: Challenge model development (Waddington *et al.* 2014); **n=88, TII.** Vaccine Randomized Control Trial: M01ZH09, Ty21a, placebo (Darton *et al.* 2016); **n=18 from VAST.** Vaccine Randomized Control Trial: Vi-TT, Vi-PS, placebo (Jin *et al.* 2017); **n=45, PATCH.** RCT: Re-challenge & Microbiome (Gibani *et al.* 2018)

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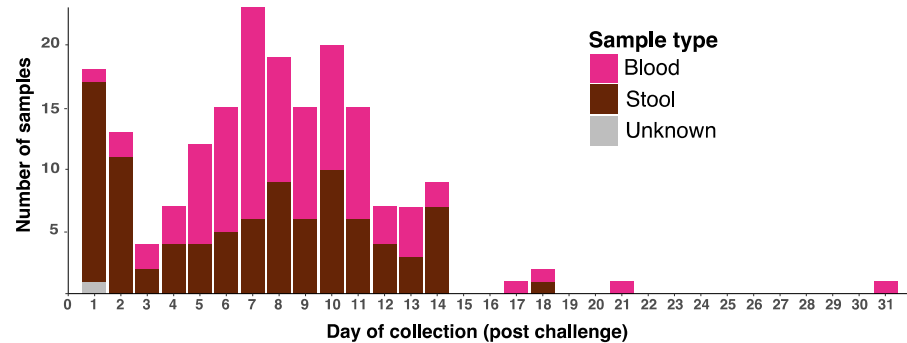
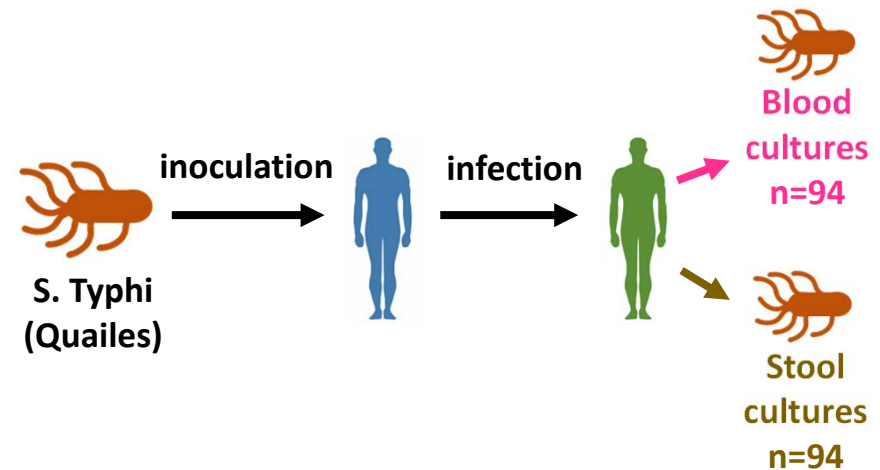
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S. Typhi *in vivo* substitution rate

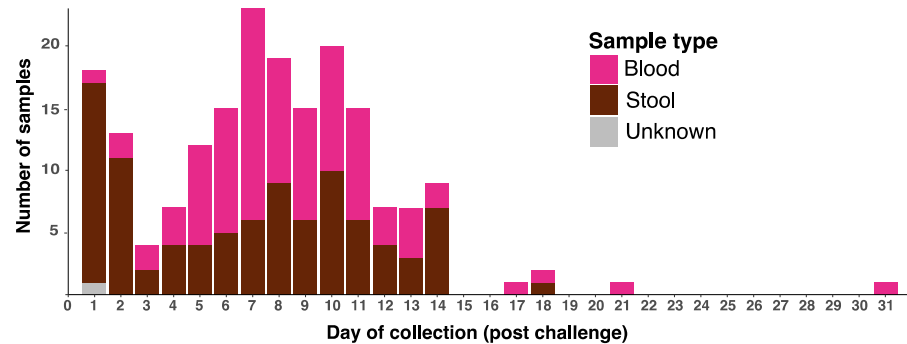
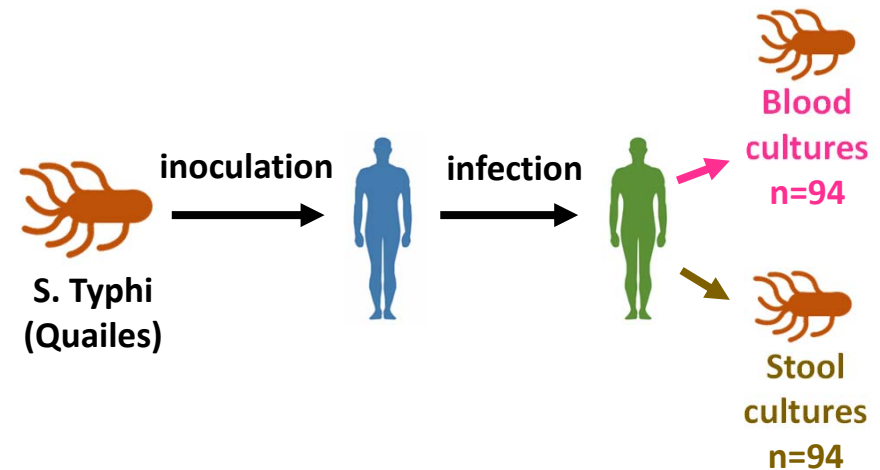
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- 0-1 SNPs per patient sample
- 7 SNPs over 1453 observed days



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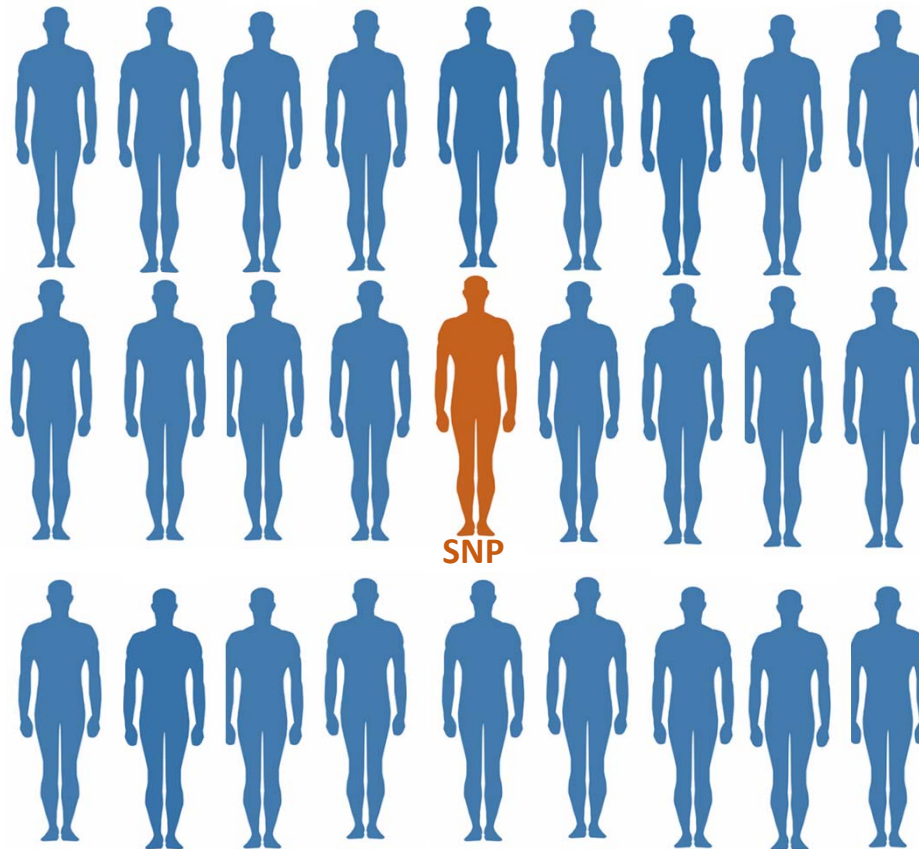
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- 189 post challenge stool and blood cultures from 4 studies
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- 7 SNPs over 1453 observed days
- 0.14 SNPs/month, 95% CI [0.069, 0.300]



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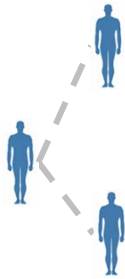


~1 in 27 samples harbored a single base change (SNP)

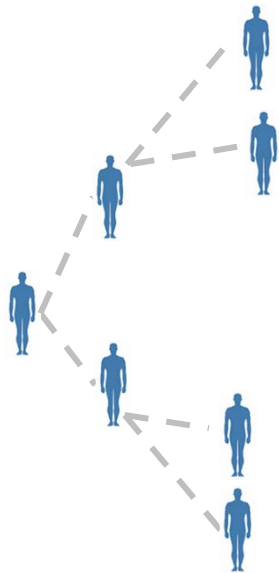
Individual transmission events



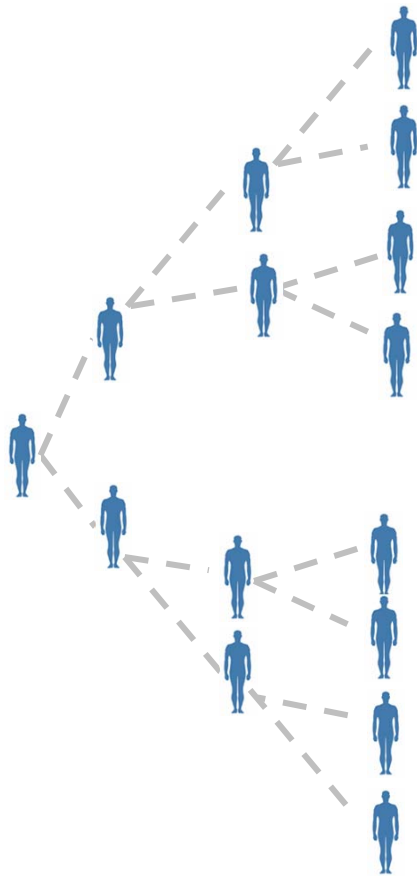
Individual transmission events



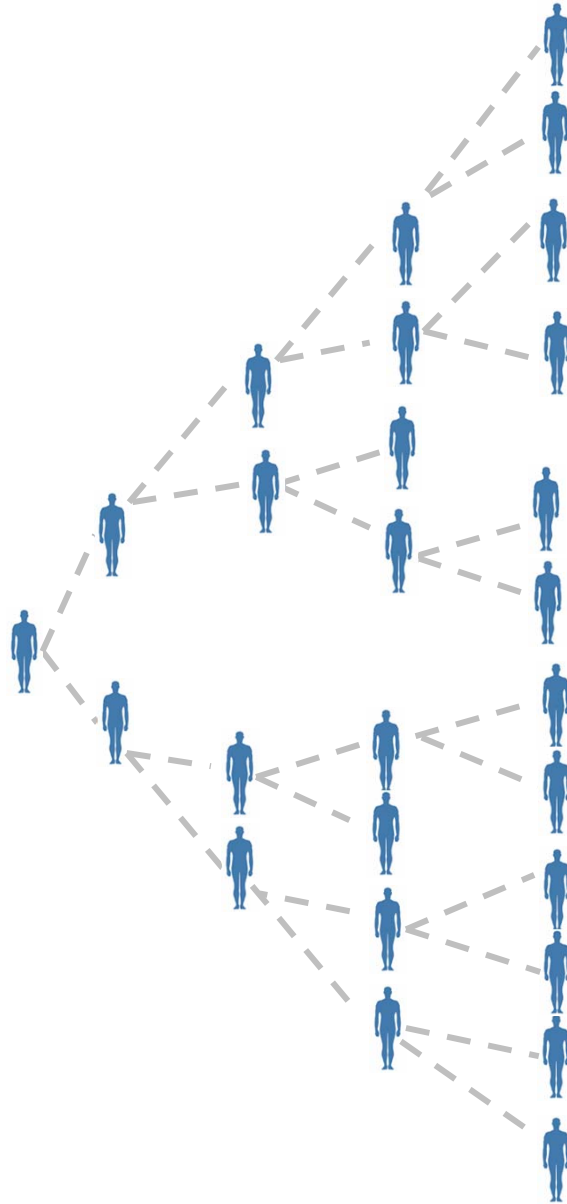
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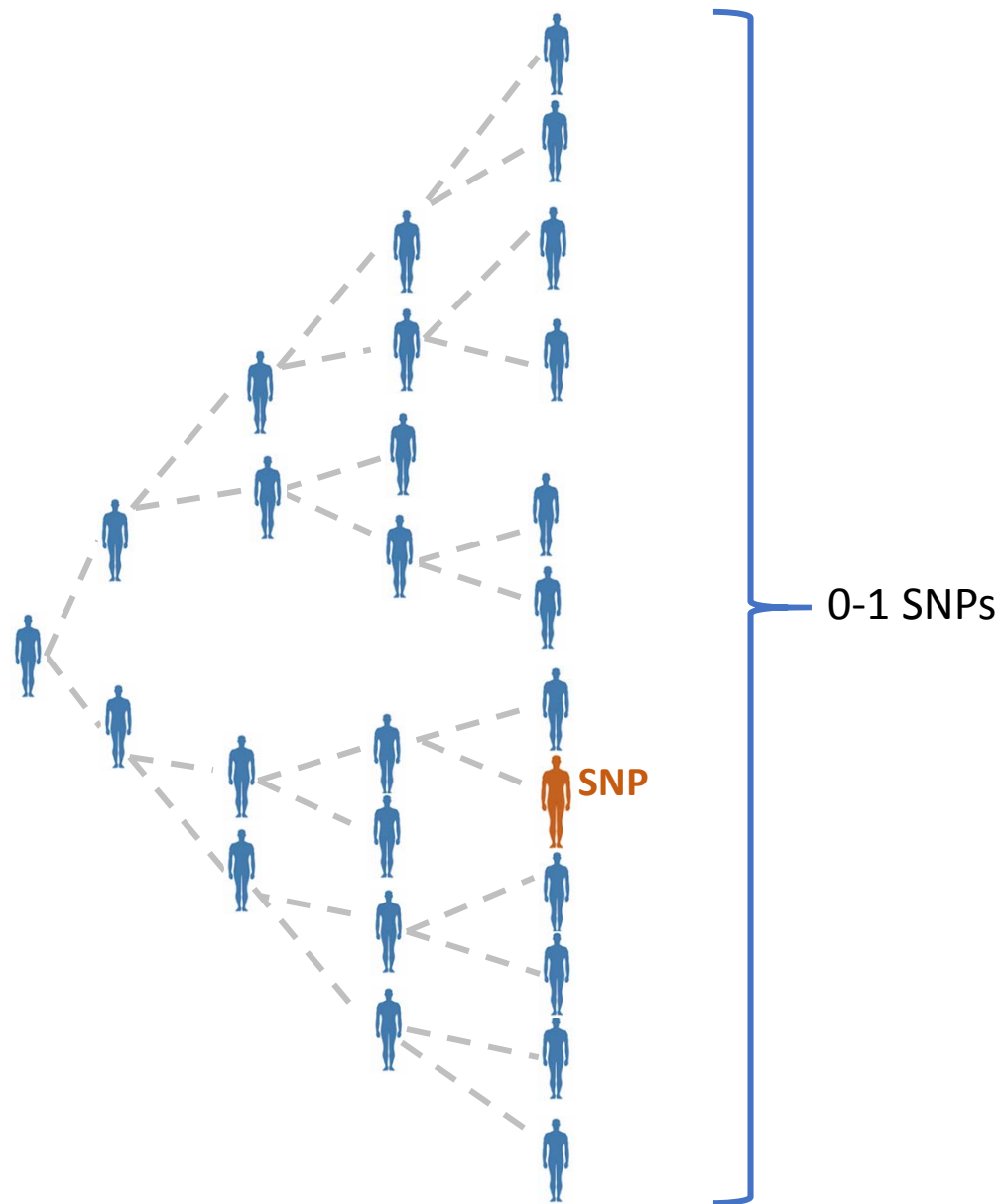
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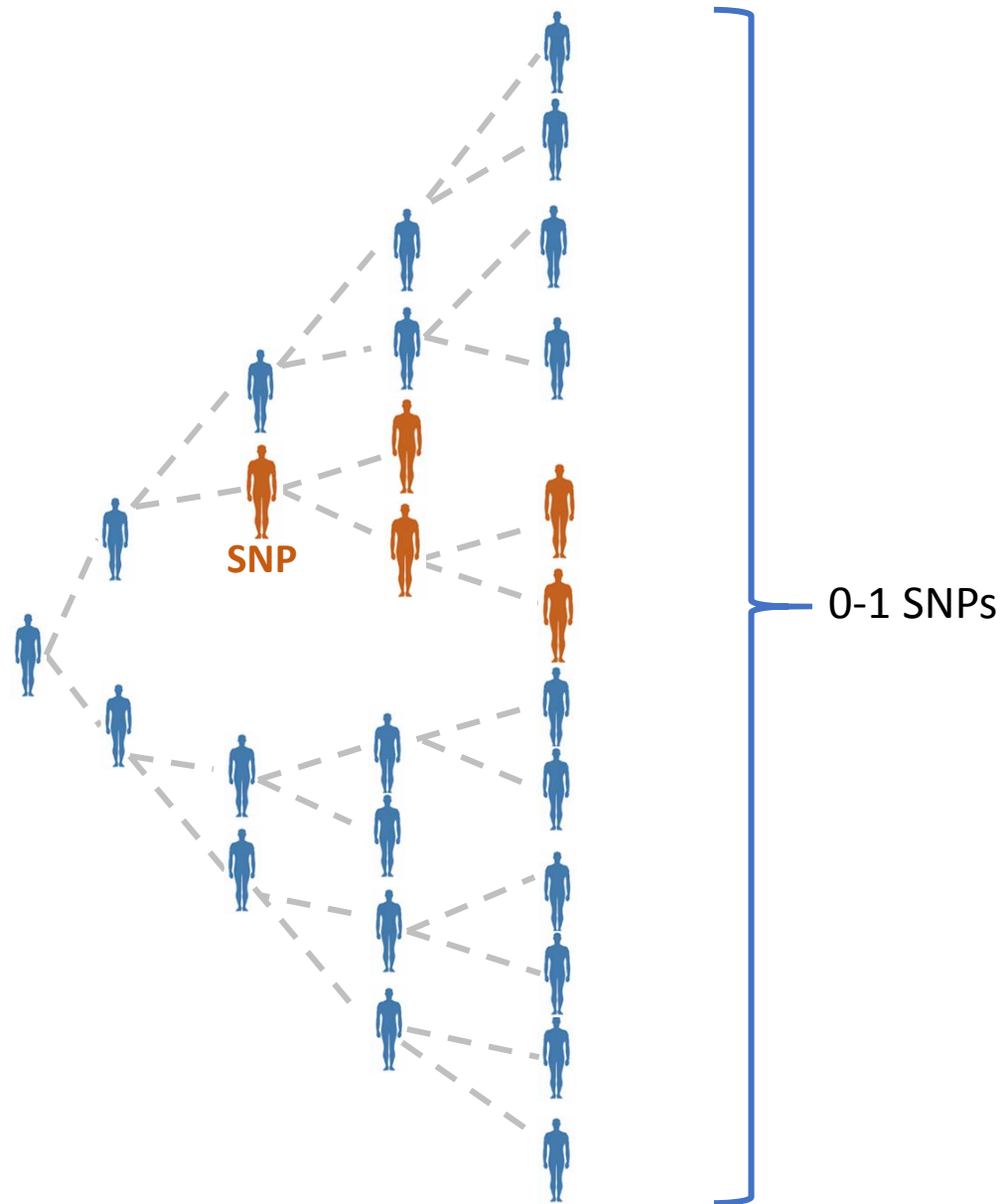
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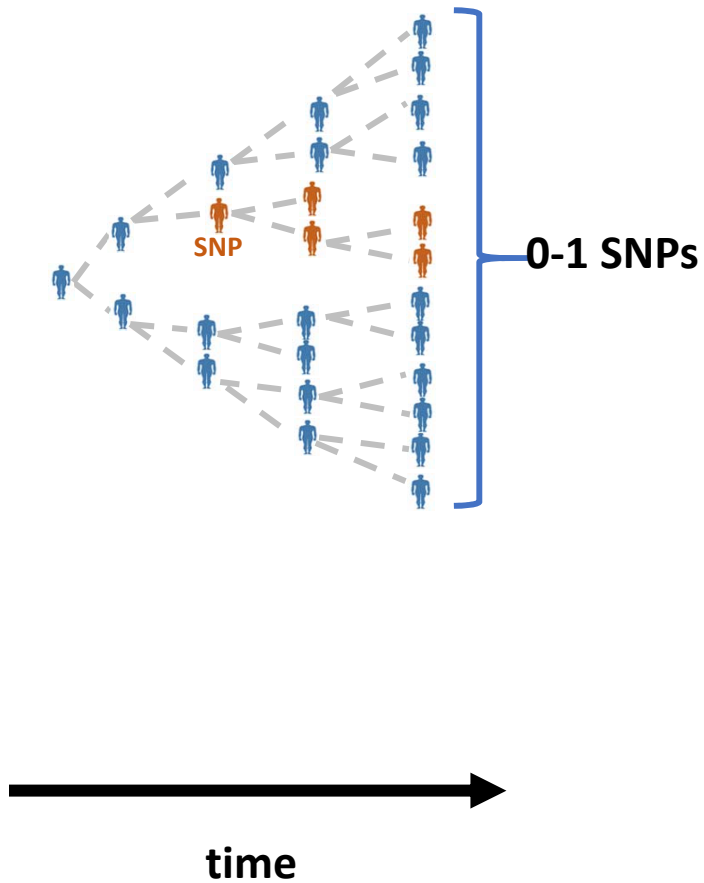
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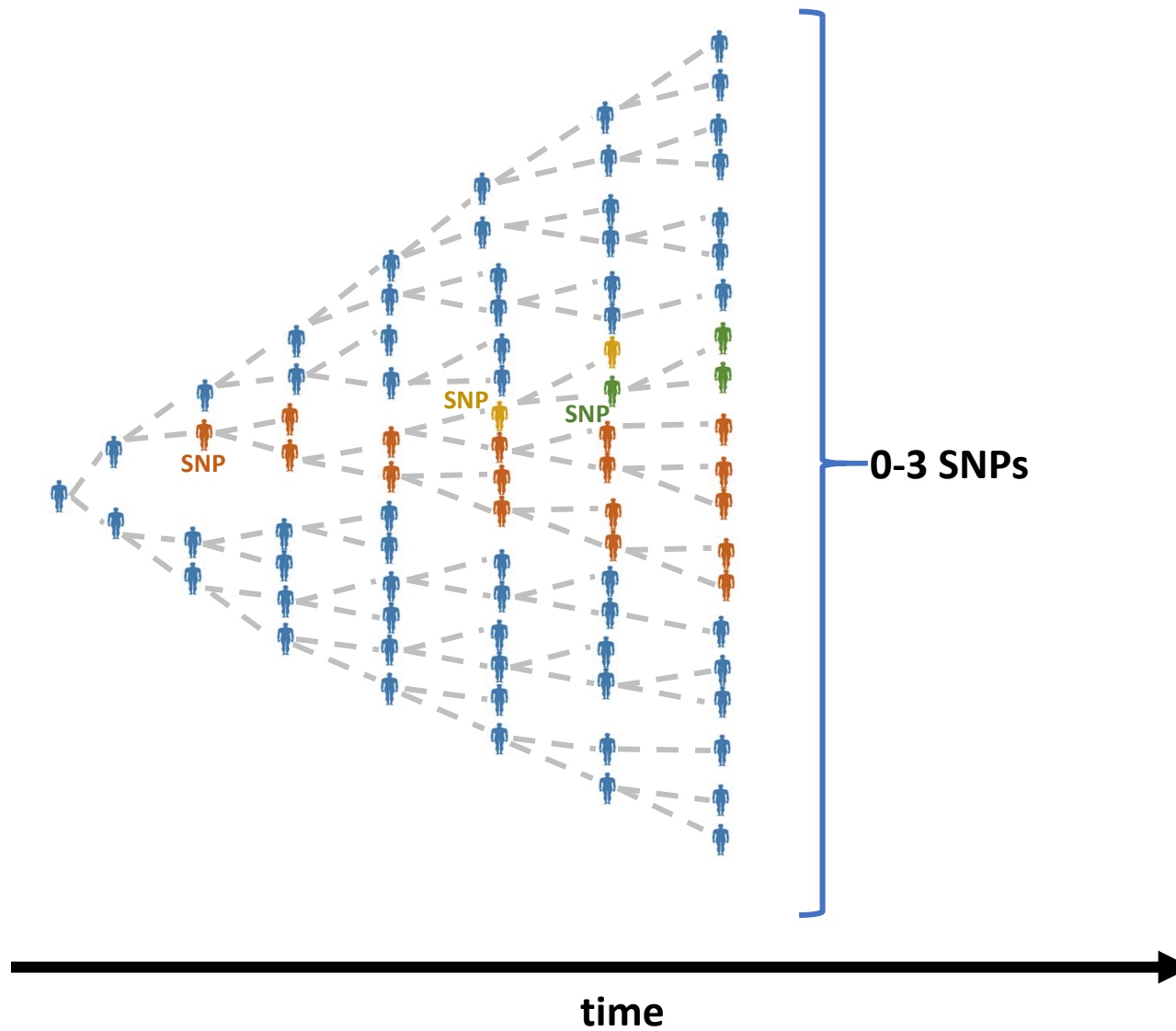
Individual transmission events



Transmission dynamics

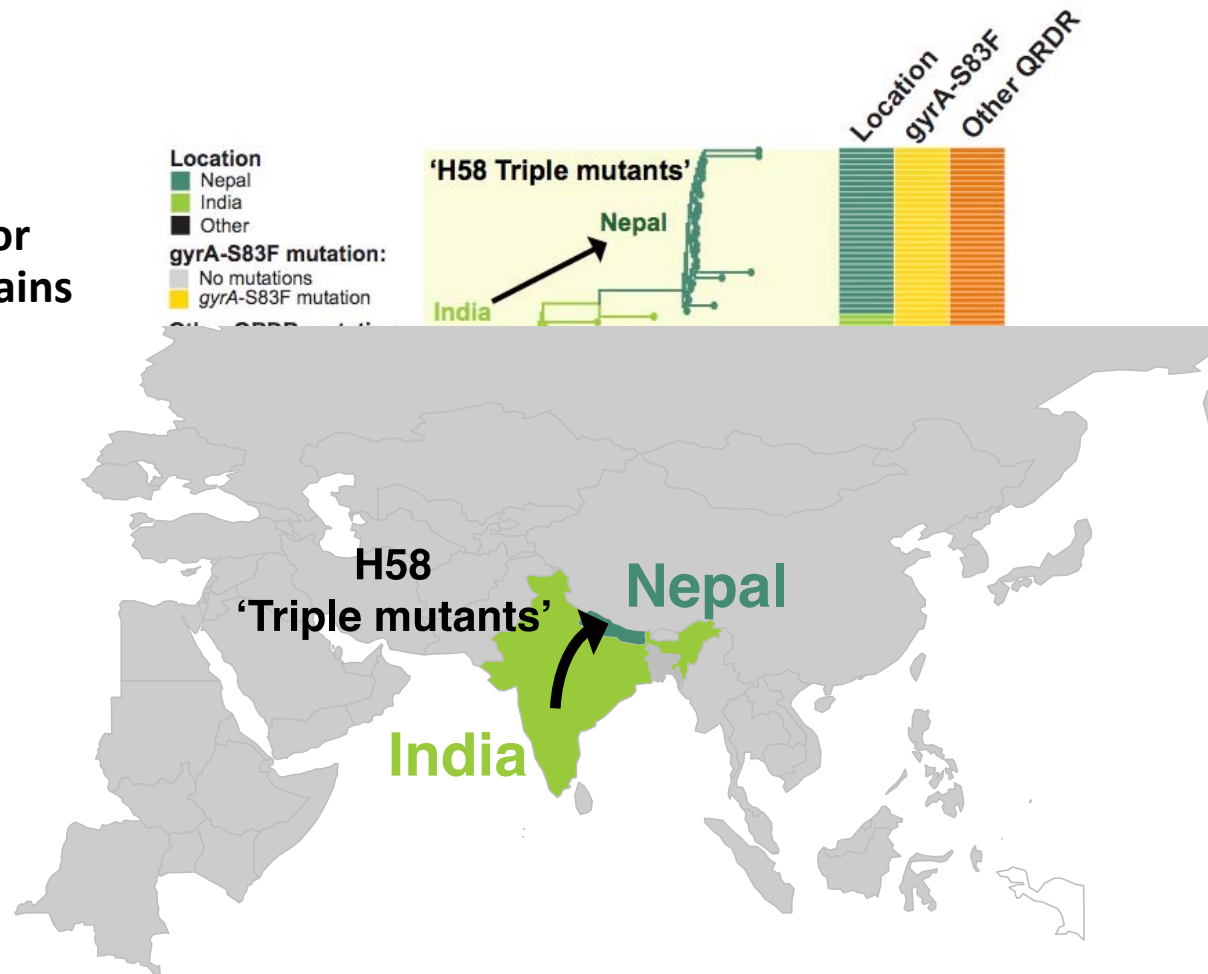


Transmission dynamics



The role of WGS in *S. Typhi* in understanding transmission dynamics

- **Classify strains by genotype**
 - e.g. H58 (4.3.1)
- **Detect Multi-drug resistant (MDR) or Extensively drug resistant (XDR) strains**
 - Detection of AMR
 - Detection of mobil elements e.g. plasmids
- **Date AMR acquisition**
- **Understand changes in structure & AMR over time**
- **Understand regional and international transmission and circulation**
- **Understanding age distribution of genotypes and other parameters**
 - e.g. seasonal patterns



Conclusions

1. Typhi evolves too slowly for WGS data to be used to understand **individual transmission** events

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1. Typhi evolves too slowly for WGS data to be used to understand **individual transmission** events
2. However, WGS data are key in understanding **transmission dynamics**, AMR, regional and global strain circulation patterns, and resolving point source outbreaks in different endemic settings

Recommendations

1. For endemic countries which do not yet have formal WGS based surveillance programmes, return traveler WGS data routinely generated by public health laboratories (e.g. PHE) are suitable for **temporary sentinel surveillance**

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1. For endemic countries which do not yet have formal WGS based surveillance programmes, return traveler WGS data routinely generated by public health laboratories (e.g. PHE) are suitable for **temporary sentinel surveillance**
2. We need to encourage **data sharing** from multiple public health laboratories, while we initiate and improve surveillance programmes within endemic areas across the world

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